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### Phytochemical, Antioxidant, Antimicrobial and Micropropagation Study of *Bambusa* Species

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#### Abstract:

Bamboo being a versatile plant provides living benefits numerous to beings comprising the environment. It is used in household works for construction, support; provides fodder for animals, edible shoots human: environment helps sequestering carbon dioxide from the atmosphere and converting the carbon into plant fiber and this benefit continues. Besides, individual plant parts such as stem, leaves possess medicinal properties which is analyzed and evaluated through various studies of phytochemical, antioxidant and antimicrobial activity. Many people are benefitted economically by selling this plant so demand for its production is escalating day by day. Micropropagation is thus considered as crucial as this ensures production throughout the year irrespective of season and hindrance caused by flowering of this plant. Among many genus of this plant, Bambusa is considered as one of the important one and has many species under this genus. This review is thus an attempt to explore studies conducted by many scholars and researchers of this genus which is intended to provide light to some of exceptional works done which eventually is beneficial to humankind.

#### Keywords

Bambusa, Phytochemical, Antioxidant, Antimicrobial, Micropropagation

#### 1. Introduction

Bamboos are perennial plants that belong to grass family Poaceae in the subfamily Bambusoideae. Woody bamboos consist of 9 to 10 sub tribes, containing between 59 and 111 genera and at least 1447 species [1]. It includes some of the fastest growing plants in the world due to its unique rhizome dependent system [2]. It does not require replanting after harvest as it will regenerate from its rhizome root structure. Besides, it improves soil quality and helps to rebuild eroded soil. The word bamboo is Kannada derived from the term 'bambu' which was found to introduced through Indonesian and Malay to English [3]. It is likely that more than 1250 species under 75 genera of bamboo are unequally distributed in the humid tropical, subtropical and temperate regions of the world [4]. It is popularly known as "poor man's timber" since it is commonly used by rural people of many countries [5]. It serves as a great relief for rural people especially of Asian countries in economical as well as social terms. It is used as building material for construction purposes and people make money by selling bamboo as a whole, handicrafts and bamboo shoot for edible purposes and many more. Bamboo is also known as the "green gold of the forest" for of its global demand and versatile uses [6]. Bamboos are present with 75 genera growing in a wide range of different habitats worldwide so there is much variation in the characteristics and behavior they exhibit. The Plant List includes 509 scientific plant

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names of species rank for the genus Bambusa out of which 130 are accepted species names. The Plant List includes a list of botanical names of plant species which was launched in 2010 and created by the Royal Botanic Gardens, Kew and the Missouri Botanical Garden. These accepted species are also included in World Checklist of Selected Plant Families (WSCP). This list further includes a further 97 scientific plant names of infraspecific rank for the genus Bambusa which is not included in WSCP. It is a genus of large clumping bamboos in the major group angiosperms (flowering plants). They are inherent to Southeast Asia, China, Taiwan, Himalayas, New Guinea, Melanesia and Northern Territory of Australia. They are also found to be acclimatized in other regions such as Africa, Latin America and different oceanic islands [7, 8]. Some members of the genus Bambusa species has been found to provide numerous benefits to science and communities [9, 10, 11]. Among twenty priority species of bamboo with economic importance which was listed in 'Priority **Species** Bamboo and Rattan', compiled by the International Network for Bamboo and Rattan (INBAR), there are seven species of Bambusa genus [12]. INBAR released a revised list that included 20 taxa (species particular economic genera) of importance and a further 18 taxa of importance [13]. Species of Bambusa genus are gaining more popularity nowadays because of various properties it possesses whether be it on construction work or better micropropagation capabilities. They also possess various antimicrobial capabilities and antioxidant activity because of different

phytochemicals they inhibit. In this review paper, phytochemical assessment, antioxidant activity, micropropagation abilities and antimicrobial activity of various species of *Bambusa* genus is being reviewed.

#### 2. Phytochemical Screening

Plants are the richest resource of drugs of traditional systems of medicine, modern nutraceuticals, medicines, food supplements, folk medicines. pharmaceutical intermediates and chemical entities for synthetic drugs [14]. Plants are capable of synthesizing a wide variety of chemical compounds that perform important biological functions and these compounds are also used to defend against attack from predators such as insects, fungi and herbivorous mammals. Photochemical can be defined as the non-nutritive chemical compounds which occur naturally in plants where 'phyto' means plant in Greek. Extraction and characterization of several active phytocompounds from plants have given rise to some high activity profile drugs [15]. Many plant extracts and phytochemicals show antioxidant or free radical scavenging properties [16]. These phytochemicals are divided into primary secondary metabolites. Primary metabolites are essential to all plants such as sugars, fats and secondary metabolites; also called phytochemicals are found in smaller range of plants and serve a specific purpose since these have protective properties.

There has been conduction of many investigations regarding study of phytochemicals on various medicinal plants from long time ago. However, study of these phytochemicals on bamboo leaves can be viewed as a novel practice and its interest is increasing more as each day passes. Bamboo leaves have therapeutic along with nutritive properties so it is a good source of fodder for livestock. Phytochemical

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investigation was performed on three bamboo species B. vulgaris Schrad. ex J. C. Wendl., B. ventricosa McClure and O. abyssinica (A. Rich.) by Coffie et.al., 2014 in order to check its safety for consumption in both the dry and wet ethanol extracted leaf samples [17]. All the leaves extract was found to contain saponin, general glycoside, coumarin and cyanogenic glycoside and extract of B. ventricosa and O. abyssinica had polyphenol and flavonoid as well. There were no traces of alkaloid, carotenoid, triterpenoid, steroid, anthraquinone and anthracene glycoside in any of the species. B. vulgaris was regarded as the safest among these three species as it contained classes of phytochemicals contained relatively less saponin as these can lyse red blood cells of larger herbivores and this is why plant extracts are not directly injected into human beings [18]. Also their study concluded that the phytochemicals present in leaves extract were not considerably affected by air drying of the leaves because of which these leaves could be fed to livestock either in the fresh state or in the dry state as hay. In the dry season where there is low rainfall, lack of fodder for livestock is still a great problem in developing countries. Bamboos can be a better alternative for fodder as their leaves are carbohydrate rich when compared with components of the leaves from several vegetables [19]. Besides, these nutritious, content of amino acid is high, has less microbial susceptibility and possess long shelf-life of their meal.

In the study performed by Joselin et.al, 2014, phytochemical analysis was done on the leaves extract of five different bamboo arundinacea (Retz) species B. Willd, B. heterostachya (Munro) Holttum. B ventricosa McClure, B. vulgaris Schrad. ex. J. C. Wendl. and *D strictus* (Roxb.) Nees [20]. Analysis was performed on six different solvents namely aqueous, petroleum ether, chloroform, ethanol and

along with Fourier acetone Transform Infrared Spectroscopy (FTIR analysis). Aqueous solvent comparatively showed better presence of phytochemicals while quinones were completely absent in all the bamboo leaf extracts. Flavonoids and coumarins were found to be present in seven extracts whereas glycosides in six extracts indicating their way for future use as antioxidant agents. The bamboo contains 2% to 5% flavone and phenolic compound that have the power to remove active free radicals, stop sub-nitrification reduce blood fat [21]. phytochemical screening of leaf extracts of B. vulgaris revealed the presence of phenolic compounds, flavonoids, terpenoids, alkaloids, tannins, alkaloids when extracted on hexane, chloroform, ethyl acetate and water solvents [22]. The plant extracts were good sources of different classes of bioactive compounds. These phytochemicals have therefore good potential demonstrated for expansion of modern chemotherapies against microbial infections as the crude extracts considerably inhibited the growth of fungi A. niger.

Not only leaves and stem, bamboo shoots of species are considered equally few important since it has suitable health benefits because of their rich contents in proteins, carbohydrates, vitamins, fibres, minerals and very low fat. Bamboo shoots contain many minerals like potassium, calcium, manganese, zinc, chromium, copper, iron and also the minor amounts of phosphorus and selenium [23], which are important for human body. Fresh bamboo shoot also possesses several vitamins such as Vitamin A, vitamin B1, vitamin B3, vitamin B6 [24]. Besides, bioactive compounds such as flavonoids, phenols and phenolic acids which are found to have potential health benefits are also found in bamboo shoot [25]. Its consumption as healthy diet is increasing rapidly mainly in

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Southeast Asian and East Asian countries [26, 27]. The most widely distributed phytosterols, sitosterols are abundantly found in fermented bamboo shoots [28]. Fermented bamboo shoots have been eaten by various tribes since ancient times [29, 30]. Likewise, dish prepared with fermented bamboo shoots is taken as local delicacy in many ethnic communities. Phytochemical screening of fermented B. balcooa shoot revealed the presence of tannins, steroids, glycosides, flavanoids, phenols, carbohydrates and proteins when done in methanol and ethyl acetate solvents [31]. More phytochemical constituents were shown by methanol solvent in comparision to ethyl acetate. In study conducted by Srivastava, 1990, the concentration of total phytosterols in dried samples of fresh shoots of B. tulda and D. giganteus was 0.21-0.39% and 1.6-2.8% in samples of fermented shoots which was determined colorimetrically [32]. Anaerobic digestion was the cause for increase in level of phytosterols in fermented shoots as it degrades the organic matter. Therefore, fermented bamboo shoots could be used as a starting material for the production of steroidal drugs which can reduce inflammation and pain.

#### 3. Antioxidant Activity

An antioxidant is broadly defined as any substance that delays or inhibits oxidative damage to a target molecule by trapping free radicals [33]. There is increment in the interest and study of antioxidant activity of various bamboo leaves among scholars and worldwide researchers although antioxidative compounds in bamboo leaves are not fully known. Many age related human diseases are also the result of cellular damage by free radicals and antioxidants could play a crucial role in preventing such diseases. The medicinal effects of bamboo leaves are mostly attributed by their

antioxidant capacity [34]. When plant derived phytochemicals, are consumed in a regular basis, Halliwell B, 1996 pointed that it may drift the balance toward an ample antioxidant status [35]. The study of phytochemicals and potential antioxidant activity of plant extract is thus important in medical world as well. Flavonoids are diverse group of phytonutrients found in nearly all fruits and vegetables, grains, bark, roots, stems, flowers, tea and wine with than 6,000 types. These responsible for providing all those vibrant colors in fruits and vegetables along with carotenoids. These have the capacity to reduce free radical formation and scavenge them so interest is growing on study of relationship of flavonoids and antioxidant activity particularly for the reason of health benefits. Besides, the antioxidant capacities of many flavonoids are much found stronger than those of vitamins C and E [36]. Many groups of flavonoids have also been used in treatment of human diseases [37]. Both the antioxidant and prooxidant effects of flavonoids are discussed by Procházková et. al. 2011 in their review paper [38]. Flavonoids cannot only be purely accounted for antioxidants as meanwhile under certain reaction conditions they can also display prooxidant activity promoting the oxidation compounds. other Nonetheless, prooxidant or antioxidant properties of a particular flavonoid depend most of all on its concentration. Flavonoids have also been stated to as 'nature's biological response modifiers as they possess inherent ability to alter the body's reaction to allergies, viruses and carcinogens [39].

Study of antioxidant activity of fifteen different bamboo species was performed by Wang et.al, 2007 among which eight species were of *Bambusa* [40]. Their results revealed that the methanolic extract of *B. textilis* McClure possessed the highest antioxidant activity among the selected fifteen bamboo species. It's

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extract was then further analysed by HPLC-UV and HPLC-micro-fractionation to identify antioxidant compounds, where three antioxidant fractions namely 4''-O-β-D-xylopyranoside, isoorientin 2"-*O*-α-L-rhamnoside isoorientin isoorientin were isolated. Evaluation of total polyphenol and antioxidant activity of leaves extract of B. nutans and B. vulgaris was carried out by Tripathi et.al, 2015 where total phenolic content of the extracts spectrophotometrically calculated using a modified Folin-Ciocalteu method and DPPH (2,2-diphenyl-1-picrylhydrazyl) radical scavenging assay was used to assess antioxidant activity [41]. Their study revealed that B. nutans can be beneficial and has higher polyphenolic content and antioxidant activity than B. vulgaris. Therefore, it can be speculated that B. nutans can also be potentially useful as a natural source of antioxidant or in medicine. The higher antioxidant and superoxide scavenging activity in the chlorogenic acid (CGA) methyl ether compounds than in other phenolic acids and antioxidants is mediated through odihydroxyl and methoxyl properties of the molecules as well as inhibition of xanthine oxidase [42]. In their study, three CGA derivatives from the leaves of bamboo P. edulis was isolated and identified as 3- O-(3'-methylcaffeoyl)quinic acid, 5- O-caffeoyl-4-methoxylquinic acid and 3- Ocaffeoyl-1- methoxylquinic acid. CGA is an ester of caffeic acid with quinic acid found naturally in various plants such as coffee beans, apples and blueberries. DPPH radical scavenging activity, total phenols and antioxidant activities was assessed and analyzed by Goyal et.al, 2010 in Indian wild B. vulgaris "Vittata" methanolic leaf extract [43]. There was found presence of carbohydrates, reducing sugars, flavonoids, steroids, saponins, alkaloids, tannins, anthraquinones and glycosides. The high contents of total

phenolic compounds and total flavonoids indicated that these compounds contributed to the antioxidative activity. Phytochemical especially flavonoid is responsible for antioxidant activity and many species of *Bambusa* exhibit this property. Further research is still necessary for more qualitative and quantitative analysis of these compounds to derive potential use of bamboo leaves in medicinal world.

#### 4. Antimicrobial Activity

In today's world, the discovery and development of antibiotics is considered as one of the most successful and powerful achievements of modern science and technology which is intended to control infectious diseases. There is increase in resistance of pathogens to traditional antimicrobial agents. This resistance has indicated in need for isolation of microbial agents which is less susceptible to regular antibiotics; also allowing the recovery of resistant isolates during antibacterial Although there is therapy. much advancement in medical world, there is still a critical need to control antimicrobial resistance by upgraded antibiotic usage in order to reduce hospital cross-infection [44]. The medicinal plants containing compounds with antimicrobial activity are found all around the world and many efforts have been made to discover novel effective antimicrobial compounds. Antimicrobial screening is done to specify the presence of antimicrobial molecule in the plant against pathogen and microbes.

Micro dilution assay showed that ethanolic extract of *D. asper* leaves extract was effective to inhibit all tested *E. coli* strains [45]. The chemical identification using pyrolysis GC/MS revealed that fatty acids, together with esters, long chain alcohols and aldehydes were the major compounds in ethanolic extract. Also, antidiarrheal agent could be extracted from *D. asper* leaves extract. In similar experiment, antibacterial

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activity of B. arundinaceae was tested the strains of S. against P.aeruginosa, E. coli and Bacillus [46]. Ethanolic extract was moderately effective against all four organisms, while aqueous extract showed moderate effect against E. coli and S. aureus. Bamboo is considered as a good source of phytosterols that are the precursors of various pharmaceutically active steroids found in plants and these also act as nutraceuticals. In the study. conducted by Wasnik and Tumane, 2014, they found that the ethanolic and methanolic leaves extract of B. bambose L. showed excellent inhibitory action against all tested ten multiple drug resistant bacteria and they all were clinical isolates from wound infection [47]. Here also ethanol solvent comparatively showed better results like previous studies. Effectiveness of plant extract against microbial strains was determined by measuring zone of inhibition (ZOI) in all the studies. Ethanolic extracts of B. arundinacea and M. indica had higher inhibition on both gram positive as well as gram negative bacteria indicating both are potential source of natural antioxidants, phytochemicals and antibacterials. [48]. M. indica showed more therapeutic capacity. In this study also, ethanolic and methanolic extracts of both the tested plants were more effective than aqueous extracts. Antimicrobial screening of the n-hexane, chloroform and ethyl acetate extracts of B. vulgaris showed that the crude extracts inhibited the growth of fungi A. niger. This supports the ethno medicinal use of aqueous extracts of this species to treat sexually transmitted diseases and for wounds [22]. However, although bamboo leaves possess antibacterial activity there is not much studies and research conducted in order to find its efficacy and usefulness till date. Further investigations are still needed to identify their vital components needed for their effective role as therapeutic plant for those already studied extracts as well.

Besides, additional work should also be carried out in order to fractionate the extracts to draw a better understanding of mechanism of those antioxidants responsible for antimicrobial activity.

#### 5. Micropropagation

It is a rapid multiplication of a selected plant by use of vitro culture techniques. This technique utilizes modern plant tissue culture methods. Explants upon subsequent subcultures result in a number of multiple shoots and these multiple shoots on elongation allow rooting in vitro. After rooting, they are in vitro hardened and transferred to field [49]. Traditionally, bamboos were propagated with seeds, clump division, culm and rhizome cuttings but due to gregarious flowering, low seed viability, high costs, problems faced during long-distance transportation of vegetative propagules and poor efficiency of plant production alternative propagation methods were introduced [50]. Micropropagation of bamboo is the only viable method [51] and it has has allowed in developing new type of ornamental or commercial bamboo. The plantlet following this method can be produced in any time of the year without depletion in its quality. This method is regarded as the best available technique and will serve as standard for mass scale propagation of bamboo in the future [52]. For tissue culture of bamboo, the use of starting material i.e. seeds or adult plants including the choice of the propagation method are crucial [53]. In comparision to other medicinal plants, tissue culture in bamboo was practiced lately. The aseptic germination of bamboo seeds in D. strictus was stated by Alexander and Rao, 1968 [54] and this is reported to have indicated the start of tissue culture of bamboo in White, 1964 major and minor elements [55]. Following this, this medium has been then used by Nadguada et al, 1990 in B. arundinacea and D. strictus; Mascarenhas

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et. al, 1988 in D. strictus; Ravikumar et. al., 1998 in D. strictus for zygotic embryo germination [56-58]. After use of this medium, Murashige and Skoog ,1962 (MS) basal medium which is more popular these days was regularly used in embryo germination of B. arundinacea where half strength of MS basal medium was used, in B. nutans and D. membranaceus and in B. tulda. MS medium is fortified with cytokinins, auxins, glucose source, gelling agent and carbon source according to the type of explant cultured as well as plant species involved [59-62]. Contamination and browning of explant is one of the main reasons of failure of tissue culture experiment. The severity of browning has varied according to species, tissue or organ, and nutrient medium and other tissue variables [63]. However, many techniques have already been evolved and studied to overcome these problems. Besides above mentioned, there are many other researchers who have accomplished successfully in propagating various bamboo species from zygotic embryo. Some of these are in B. arundinacea in Nitsch and Nitsch medium. 1967; in MS medium; in B. vulgaris in MS medium [64-67]. Various plant parts such as nodal segments, axillary buds were then used as explants as this ensured large scale propagation of bamboo. Micropropagation through axillary bud proliferation by nodal explants did not interfere with the production of true to type plant. This method also does not interrupt the callus phase and thus maintains the clonal fidelity [51]. This purpose of micropropagation was well satisfied by MS media so almost all researchers relied on using this medium as this media supplemented all the essential nutrients required for development of new plantlet invitro. Nodal explants were used by Arya et. al for B. arundinacea and B. vulgaris, Das and Pal and Islam and Rahman, for B. balcooa, Arya and Sharma for B. bamboos, Lin et.al for B. edulis,

Shirin and Rana for *B. glaucescens*, Islam and Rahman and Yasodha et. al for *B. nutans*, Mishra et. al for *B. tulda*, Ndiaye et.al for *B. vulgaris* [68-76]. Likewise, axillary shoots was used as explant by for *B. atra* and axillary bud for *B. vulgaris* 'striata' [77-78].

Nodal explant is considered as the best starting material for micropropagation where multiple shoots are preferred or required as this is capable of providing An efficient micropropagation such. multiplication protocol for balcooa from nodal explants was established where 88% success rate of acclimatization was achieved when done in liquid MS media [79]. In the same species, Patel et.al achieved survival rate of 74.66% when nodal segments were cultured on both solid and liquid MS media, Sharma and Sarma, achieved 100% success in field transfer, 90% success rate achieved by Mudoi and Borthakur whereas Ansari et. al achieved 90% success rate when culture was initiated in solid MS media [80-83]. In similar experiment, Raju and Roy established an efficient and reproducible micropropagation protocol for mass production and generation of uniform clones for B. bambos (L.) Voss where explant used was nodal segments containing unsprouted axillary buds in liquid MS media [84]. They recorded 100% survivability from wellestablished rooted plantlets during acclimatization process.

B. tulda is another species which researchers show great interest to mass propagate as its main purpose is on construction works and has high demand. An efficient and rapid method of in vitro propagation of this species from aseptic seedlings was studied and achieved 90% success rate upon field transfer when shoots from 3-week-old seedlings in liquid medium were used to initiate the cultures [85] whereas Mishra et.al had 98% rooting in liquid MS media [86]. In same species,

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Waikhom and Louis had 81.81% success when culture was initiated on liquid MS media [87]. They take the privilege of successfully developing protocol which could produce rhizome in half-strength MS the first medium for time of species. Survival rate was 100% when semisolid MS medium was used for culture of nodal explant [88]. B. vulgaris Schrad. ex Wendl has been promoted in order to solve the deforested environments and economic problems [89]. During their study they attained 75% survival where their explant was inter-nodal region of stem and culture medium was solid MS media. B. nutans is edible ornamental bamboo and is also propagated in vitro by many researchers for both study and commercial purpose. 95% survival rate was obtained by Mudoi et. al whereas 100% success rate was observed by Sharma and Sarma on field transfer when nodal explants were used in MS medium [90,91]. Research on other species of this genus is also being carried out which can ultimately contribute in mass production of bamboo.

#### 6. Conclusion

There have been many developments in research techniques and equipment which has allowed for the assessment and evaluation of many constituents of plant molecule and same applies for bamboo plant also. It is undoubtedly a versatile plant and can serve to wellbeing of living beings in various ways. There has been conduction of lot of works to find the beneficial active constituents of some species, but still there is a long way to go to assess the true potential of all the species of this plant.

#### 7. References

[1] Ohrnberger, D. (1999). The bamboos of the world: annotated nomenclature and literature of the species and the higher and lower taxa. Elsevier.

- [2] *Illustrated Oxford Dictionary*. Great Britain: Oxford University Press. 1998. p. 69. ISBN 140532029-X.
- [3] Farrelly, David (1984). The Book of Bamboo. Sierra Club Books. ISBN 0-87156-825-X.
- [4] Rai, P. K. (2009). Comparative assessment of soil properties after bamboo flowering and death in a tropical forest of Indo-Burma hot spot. *AMBIO: A Journal of the Human Environment*, 38(2), 118-120.
- [5] Keshari, V. P. (2005). Bamboo: From poor man's timber to green gold. *Hamro Kalpana Brikshya*, 164, 10-4.
- [6] Sharma, Y. M. L. (1980). Bamboos in the Asia Pacific Region. In *Bamboo research in Asia:* proceedings of a workshop held in Singapore, 28-30 May 1980. IDRC, Ottawa, ON, CA.
- [7] Chapman, G. P. (1997). The bamboos. In *Linnean Society Symposium Series* (No. 19).
- [8] Clayton, W. D., & Renvoize, S. A. (1986). Genera graminum. Grasses of the world. *Genera* graminum. Grasses of the World., 13.
- [9] Tripathi, Y. C. (1998). Food and nutrition potential of bamboo. *MFP News*, 8(1), 10-11.
- [10] Bao, J. (2006). The nutrition and bioactive function of bamboo shoots. *Food Nut in China*, 4, 2-3.
- [11] Chongtham, N., Bisht, M. S., & Haorongbam, S. (2011). Nutritional properties of bamboo shoots: potential and prospects for utilization as a health food. *Comprehensive Reviews in Food Science and Food Safety*, 10(3), 153-168.
- [12] Benton, A. (2015). Priority species of bamboo. In *Bamboo* (pp. 31-41). Springer, Cham.
- [13] Rao, A. N., Ramanatha Rao, V., & Williams, J.T. (1998). Priority species of bamboo and rattan.
- [14] Hammer, K. A., Carson, C. F., & Riley, T. V. (1999). Antimicrobial activity of essential oils and other plant extracts. *Journal of applied microbiology*, 86(6), 985-990.
- [15] Mandal, V., Mohan, Y., & Hemalatha, S. (2007). Microwave assisted extraction—an innovative and promising extraction tool for medicinal plant research. *Pharmacognosy reviews*, *1*(1), 7-18.
- [16] Larson, R. A. (1988). The antioxidants of higher plants. *Phytochemistry*, 27(4), 969-978.
- [17] Parekh, J., & Chanda, S. (2007). In vitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turkish Journal of Biology*, 31(1), 53-58.
- [18] Coffie, G. Y., Antwi-Boasiako, C., & Darkwa, N. A. (2014). Phytochemical constituents of the leaves of three bamboo (Poaceae) species in Ghana. *Journal of Pharmacognosy and Phytochemistry*, 2(6).

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#### **International Journal of Research**

Available at https://pen2print.org/index.php/ijr/

- [19] Tyler, V.E., Brady, L.R., & Robbers, J.E. (1995). Pharmacognosy, Lea and Fibiger, PA, USA, 57-58
- [20] Antwi-Boasiako, C., Coffie, G. Y., & Darkwa, N. A. (2011). Proximate composition of the leaves of *Bambusa* ventricosa, Oxytenanthera abyssinica and two varieties of *Bambusa* vulgaris.
- [21] Joselin, J., Jenitha, S., Brintha, T. S. S., Jeeva, S., Sukumaran, S., & Geetha, V. S. (2014). Phytochemical and FT-IR Spectral analysis of certain Bamboo species of South India. J Biodivers Biopros, 1(1), 1-9.
- [22] Ogunjinmi, A. A., Ijeomah, H. M., & Aiyeloja, A. A. (2009). Socio-economic importance of bamboo (Bambusa vulgaris) in Borgu local government area of Niger State, Nigeria. Journal of Sustainable Development in Africa, 10(4), 284-289.
- [23] Owolabi, M. S., & Lajide, L. (2015). Preliminary phytochemical screening and antimicrobial activity of crude extracts of Bambusa vulgaris Schrad. Ex JC Wendl.(Poaceae) from southwestern Nigeria. American Journal of Essential Oils and Natural Products, 3(1), 42-45.
- [24] Pandey, A. K., Ojha, V., & Choubey, S. K. (2012). Development and Shelf-life Evaluation of Value Added Edible. American Journal of Food Technology, 7(3), 363-371.
- [25] Singhal, P., Bal, L. M., Satya, S., Sudhakar, P., & Naik, S. N. (2013). Bamboo shoots: a novel source of nutrition and medicine. Critical reviews in food science and nutrition, 53(5), 517-534.
- [26] Gupta, V. K., Rachna, K., Munish, G., & Monika, G. (2010). Recent updates on free radicals scavenging flavonoids: an overview. Asian Journal of Plant Sciences, 9(3), 108-117.
- [27] Nongdam, P., & Tikendra, L. (2014). The nutritional facts of bamboo shoots and their usage as important traditional foods of Northeast India. International scholarly research notices, 2014.
- [28] Shi, Q. T., & Yang, K. S. (1992, December). Study on relationship between nutrients in bamboo shoots and human health. In Bamboo and its use. Proceedings of the international symposium on industrial use of bamboo (pp. 338-346).
- [29] Sarangthem, K., & Singh, T. N. (2003).

  Biosynthesis of succulent bamboo shoots of Bambusa balcooa into phytosterols and its biotransformation into ADD. Acta Botanica Sinica, 45(1), 114-117.

- [30] Giri, S. S., & Janmejay, L. S. (1994). Changes in soluble sugars and other constituents of bamboo shoots in soibum fermentation. Journal of food science and technology. Mysore, 31(6), 500-502.
- [31] Tamang, J. P., & Sarkar, P. K. (1996). Microbiology of mesu, a traditional fermented bamboo shoot product. International journal of food microbiology, 29(1), 49-58.
- [32] Singh, S. A., Bora, T. C., & Singh, N. R. (2012).

  Preliminary phytochemical analysis and antimicrobial potential of fermented Bambusa balcooa shoots. The Bioscan, 7(3), 391-394.
- [33] Srivastava, R. C. (1990). Bamboo, new raw material for phytosterols. Current Science, 59(24), 1333-1334.
- [34] Yamagishi, S. I., & Matsui, T. (2011). Nitric oxide, a janus-faced therapeutic target for diabetic microangiopathy—Friend or foe?. Pharmacological research, 64(3), 187-194.
- [35] Van Hoyweghen, L., Karalic, I., Van Calenbergh, S., Deforce, D., & Heyerick, A. (2010). Antioxidant flavone glycosides from the leaves of Fargesia robusta. Journal of natural products, 73(9), 1573-1577.
- [36] Halliwell, B. (1996). Commentary oxidative stress, nutrition and health. Experimental strategies for optimization of nutritional antioxidant intake in humans. Free radical research, 25(1), 57-74.
- [37] Frei, B., & Higdon, J. V. (2003). Antioxidant activity of tea polyphenols in vivo: evidence from animal studies. The Journal of nutrition, 133(10), 3275S-3284S.
- [38] Cushnie, T. T., & Lamb, A. J. (2005).

  Antimicrobial activity of flavonoids. International journal of antimicrobial agents, 26(5), 343-356.
- [39] Procházková, D., Boušová, I., & Wilhelmová, N. (2011). Antioxidant and prooxidant properties of flavonoids. Fitoterapia, 82(4), 513-523.
- [40] Huang, C., Chen, Y., Zhou, T., & Chen, G. (2009). Sulfation of dietary flavonoids by human sulfotransferases. Xenobiotica, 39(4), 312-322.
- [41] Wang, J., Yue, Y. D., Tang, F., & Sun, J. (2012). TLC screening for antioxidant activity of extracts from fifteen bamboo species and identification of antioxidant flavone glycosides from leaves of Bambusa. textilis McClure. Molecules, 17(10), 12297-12311.
- [42] Tripathi, Y. C., Jhumka, Z., & Anjum, N. (2015).

  Evaluation of total polyphenol and antioxidant activity of leaves of Bambusa nutans and Bambusa vulgaris. Journal of Pharmacy Research, 9(4), 271-77.

### (R)

#### International Journal of Research

Available at https://pen2print.org/index.php/ijr/

- [43] Kweon, M. H., Hwang, H. J., & Sung, H. C. (2001). Identification and antioxidant activity of novel chlorogenic acid derivatives from bamboo (Phyllostachys edulis). Journal of agricultural and food chemistry, 49(10), 4646-4655.
- [44] Goyal, A., Middha, S., & Sen, A. (2010). Evaluation of the DPPH radical scavenging activity, total phenols and antioxidant activities in Indian wild Bambusa vulgaris" Vittata" methanolic leaf extract. Journal of Natural Pharmaceuticals, 1(1), 40-40.
- [45] Voravuthikunchai, S. P., & Kitpipit, L. (2005). Activity of medicinal plant extracts against hospital isolates of methicillin-resistant Staphylococcus aureus. Clinical Microbiology and Infection, 11(6), 510-512.
- [46] Mulyono, N. (2012). Antibacterial Activity of Petung Bamboo Dendrocalamus Asper Leaf Extract Against Pathogenic Escherichia coli and Their Chemical Identification. International Journal of Pharmaceutical & Biological Archive, 3(4).
- [47] Singh, V. K., Rahul, S., Satish, V., Shankul, K., Sumit, G., & Ashutosh, M. (2010). Antibacterial activity of leaves of BAMBOO. International journal of Pharma and bio sciences, 1(2).
- [48] Wasnik, D. D., & Tumane, P. M. (2014). Antibacterial activity of Bambusa bambose L. against multiple drug resistant bacteria isolated from clinical specimen. Int. J. Pharm. Sci. Rev. Res, 25, 215-218.
- [49] Kaur, H., Kaur, S., & B., M. (2015).

  Phytochemical, Antioxidant and Antibacterial

  Studies on Bambusa arundinacea and Magnifera

  indica. International Journal Of Pure and
  Applied Bioscience, 3(3), 87-93.
- [50] Singh A (2015) Micropropagation of Plants.

  In: Bir Bahadur et al. (eds) Plant Biology and
  Biotechnology: Volume II: Plant Genomics and
  Biotechnology, Springer New Delhi Heidelberg
  New York Dordrecht London.
- [51] Peeters, H., Gillis, K., Oprins, J., Debergh, P. C., & Gielis, J. (2001, July). Tissue culture strategies for genetic improvement of bamboo. In XX International Eucarpia Symposium, Section Ornamentals, Strategies for New Ornamentals-Part 1 552 (pp. 195-204).
- [52] Mudoi, K. D., Saikia, S. P., Goswami, A., Gogoi, A., Bora, D., & Borthakur, M. (2013).

  Micropropagation of important bamboos: a review. African Journal of Biotechnology, 12(20).
- [53] Gielis, J., & Oprins, J. (2002). Micropropagation of temperate and tropical woody bamboos from biotechnological dream to commercial reality.

- In Bamboo for sustainable development.

  Proceedings of the Vth International Bamboo
  Congress and the VIth International Bamboo
  Workshop, San Jose, Costa Rica (pp. 333-344).
- [54] Gielis, J., Goetghebeur, P., & Debergh, P. (1999). Physiological aspects and experimental reversion of flowering in Fargesia murieliae (Poaceae, Bambusoideae). Systematics and Geography of Plants, 147-158.
- [55] Alexander, M. P., & Rao, T. R. (1968). In vitro culture of bamboo embryos. Current Science, 37(14), 415-415.
- [56] White, P. R. (1964). The cultivation of animal and plant cells. Soil Science, 97(1), 74.
- [57] Nadgauda, R. S., Parasharami, V. A., & Mascarenhas, A. F. (1990). Precocious flowering and seeding behaviour in tissue-cultured bamboos. Nature, 344(6264), 335.
- [58] Mascarenhas, A. F., Nadgir, A. L., Thengane, S. R., Phadke, C. H., Khuspe, S. S., Shirgurkar, M. V., ... & Nadgauda, R. S. (1988, November). Potential application of tissue culture for propagation of Dendrocalamus strictus. In Bamboos, Current Research, Proc. Intl. Bamboo Workshop, Cochin, India (pp. 159-166).
- [59] Ravikumar, R., Ananthakrishnan, G., Kathiravan, K., & Ganapathi, A. (1998). In vitro shoot propagation of Dendrocalamus strictus nees. Plant cell, tissue and organ culture, 52(3), 189-192.
- [60] Murashige, T., & Skoog, F. (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiologia plantarum, 15(3), 473-497.
- [61] Nadgauda, R. S., John, C. K., Parasharami, V. A., Joshi, M. S., & Mascarenhas, A. F. (1997). A comparison of in vitro with in vivo flowering in bamboo: Bambusa arundinacea. Plant cell, tissue and organ culture, 48(3), 181-188.
- [62] Saxena, S. (1990). In vitro propagation of the bamboo (Bambusa tulda Roxb.) through shoot proliferation. Plant cell reports, 9(8), 431-434.
- [63] Yasodha, R., Sumathi, R., Malliga, P., & Gurumurthi, K. (1997). Genetic enhancement and mass production of quality propagales of Bambusa nutans and Dendrocalamus membranaceous. Indian Forester, 123(4), 303-306.
- [64] Huang, L. C., Lee, Y. L., Huang, B. L., Kuo, C. I., & Shaw, J. F. (2002). High polyphenol oxidase activity and low titratable acidity in browning bamboo tissue culture. In Vitro Cellular & Developmental Biology-Plant, 38(4), 358.

#### International Journal of Research

Available at https://pen2print.org/index.php/ijr/

- [65] Mehta, U., Ramanuja Rao, I. V., & Mohan Ram, H. Y. (1982). Somatic embryogenesis in bamboo. In Plant tissue culture 1982: proceedings, 5th International Congress of Plant Tissue and Cell Culture held at Tokyo and Lake Yamanake, Japan, July 11-16, 1982/edited by Akio Fujiwara. Tokyo: Japanese Association for Plant Tissue Culture,[1982?].
- [66] Nitsch, J. P., & Nitsch, C. (1969). Haploid plants from pollen grains. Science, 163(3862), 85-87.
- [67] Nadgir, A. L., Phadke, C. H., Gupta, P. K., Parsharami, V. A., Nair, S., & Mascarenhas, A. F. (1984). Rapid multiplication of bamboo by tissue culture. Silvae Genetica.
- [68] Rout, G. R., & Das, P. (1994). Somatic embryogenesis and in vitro flowering of 3 species of bamboo. Plant cell reports, 13(12), 683-686.
- [69] Arya, I. D., Rana, P. K., Satsangi, R., Muzaffar, F. S., Sharma, S., & Arya, S. (2002). Rapid and mass multiplication of bamboos through tissue culture techniques. Role of Plant Tissue Culture in Biodiversity Conservation and Economic Development, 29-39.
- [70] Das, M., & Pal, A. (2005). In vitro regeneration of Bambusa balcooa Roxb.: factors affecting changes of morphogenetic competence in the axillary buds. Plant cell, tissue and organ culture, 81(1), 109-112.
- [71] Islam, S. A. M. N., & Rahman, M. M. (2005). Micro-cloning in commercially important six bamboo species for mass propagation and at a large scale cultivation. Plant Tissue Cult Biotech, 15, 103-111.
- [72] Arya, S., & Sharma, S. (1998).Micropropagation technology of Bambusa bambos through shoot proliferation. Indian Forester, 124(9), 725-731.
- [73] Lin, C. S., Lin, C. C., & Chang, W. C. (2004). Effect of thidiazuron on vegetative tissue-derived somatic embryogenesis and flowering of bamboo Bambusa edulis. Plant Cell, Tissue and Organ Culture, 76(1), 75-82.
- [74] Shirin, F., & Rana, P. K. (2007). In vitro plantlet regeneration from nodal explants of field-grown culms in Bambusa glaucescens Willd. Plant Biotechnology Reports, 1(3), 141-147.
- [75] Yasodha, R., Kamala, S., Kumar, S. A., Kumar, P. D., & Kalaiarasi, K. (2008). Effect of glucose on in vitro rooting of mature plants of Bambusa nutans. Scientia Horticulturae, 116(1), 113-116.
- [76] Mishra, Y., Patel, P. K., Yadav, S., Shirin, F., & Ansari, S. A. (2008). A micropropagation system for cloning of Bambusa tulda Roxb. Scientia Horticulturae, 115(3), 315-318.

- Ndiaye, A., Diallo, M. S., Niang, D., & [77] Gassama-Dia, Y. K. (2006). In regeneration of adult trees of Bambusa vulgaris. African Journal ofBiotechnology, 5(13).
- [78] Ramanayake, S. M. S. D., Maddegoda, K. M. M. N., Vitharana, M. C., & Chaturani, G. D. G. (2008). Root induction in three species of bamboo with different rooting abilities. Scientia Horticulturae, 118(3), 270-273.
- [79] Ramanayake, S. M. S. D., Meemaduma, V. N., & Weerawardene, T. E. (2006). In vitro shoot proliferation and enhancement of rooting for the large-scale propagation of yellow bamboo (Bambusa vulgaris 'Striata'). Scientia Horticulturae, 110(1), 109-113.
- [80] Negi, D., & Saxena, S. (2011).Micropropagation of Bambusa balcooa Roxb. through axillary shoot proliferation. In Vitro Cellular & Developmental Biology-Plant, 47(5), 604-610.
- [81] Patel, B., Gami, B., Patel, N., & Bariya, V. (2015).One step pre-hardening micropropagation of Bambusa balcooa Roxb. Journal of Phytology, 7, 1-9.
- [82] Sharma, P., & Sarma, K. P. (2011, December). In vitro propagation of Bambusa balcooa for a better environment. In International Conference Advances inBiotechnology Pharmaceutical Sciences (pp. 248-252).
- Ansari, I., Gupta, S., Soni, S., & Baig, A. (2017). [83] In-vitro Regeneration of Bambusa balcooa (Bamboo) through Nodal Segments. Int. J. Curr. Microbiol. App. Sci, 6(10), 4901-4905.
- [84] Mudoi, K. D., & Borthakur, M. (2009). In vitro micropropagation of Bambusa balcooa Roxb. through nodal explants from field-grown culms and scope for upscaling. Current science, 962-
- [85] Raju, R. I., & Roy, S. K. (2017). Mass propagation of Bambusa bambos (L.) Voss through in vitro culture. Jahangirnagar University Journal of Biological Sciences, 5(2), 15-26.
- Saxena, S. (1990). In vitro propagation of the [86] bamboo (Bambusa tulda Roxb.) through shoot proliferation. Plant cell reports, 9(8), 431-434.
- [87] Mishra, Y., Patel, P. K., Yadav, S., Shirin, F., & Ansari, S. A. (2008). A micropropagation system for cloning of Bambusa tulda Roxb. Scientia Horticulturae, 115(3), 315-318.
- [88] Waikhom, S. D., & Louis, B. (2014). An effective protocol for micropropagation of edible bamboo species (Bambusa tulda and Melocanna



#### **International Journal of Research**

Available at https://pen2print.org/index.php/ijr/

- baccifera) through nodal culture. The Scientific World Journal, 2014.
- [89] Pratibha, S., & Sarma, K. P. (2013). In vitro propagation of Bambusa tulda: an important plant for better environment. Journal of Environmental Research And Development Vol, 7(3).
- [90] Kaladhar, D., Tiwari, P. and Duppala, S. (2017).

  A Rapid in vitro Micro Propagation of Bambusa
  Vulgaris Using Inter-Node Explant.

  International Journal of Life- Sciences Scientific
  Research, 3(3).
- [91] Mudoi, K. D., Saikia, S. P., & Borthakur, M. (2014). Effect of nodal positions, seasonal variations, shoot clump and growth regulators on micropropagation of commercially important bamboo, Bambusa nutans Wall. ex. Munro. African Journal of Biotechnology, 13(19).
- [92] Pratibha, S., & Sarma, K. P. (2014). In vitro propagation of Bambusa nutan in commercial scale in assam, India. Journal of Environmental Research and Development, 9(2), 348.