

Determination of Mineral Content and Estimation of Total Flavonoid Content of *Brassica oleracea* L.var. italica plenck.

Arnt Win¹, Aye Mon Thida Nyo², Tin Zar Hlaing³, Khin Thu Zar³

¹Associate professor, Department of Chemistry, Kyaukse University, Myanmar ²Associate professor, Department of Chemistry, University of Mandalay, Myanmar ³Lecturer, Department of Chemistry, Kyaukse University, Myanmar

Abstract: In this research work, the Brassica oleracea L.var. italica plenck., Myanmar name Pan Gaw Bi Sein, Broccoli, which is one of the most commonly consumed vegetables by the Myanmar population was selected to qualify and quantify the flavonoids present in it. The fresh broccoli was purchased from the local market, Chan Mya Thar Si Township, Mandalay Region, Myanmar. Firstly, the mineral contents of selected sample were measured by EDXRF method. Moreover, the fresh broccolis were crushed to obtain the expressed juice with 95% EtOH which is the liquid product. This extract was checked for qualitative test of flavonoids. In addition, total flavonoid content of B. oleracea L.var. italica plenck. was evaluated by the aluminum chloride (AlCl₃) method using UV spectrophotometer (UV-1800, SHIMADZU, UV spectrophotometer) at 415 nm. The total flavonoid content of this selected sample was determined as 10.17 ± 0.16 mg quercetin equivalent (QE) per 100 g fresh weight.

Keywords : *Brassica oleracea* L.var. italica plenck., EDXRF method, UV spectrophotometer, flavonoids, quercetin.

1. Introduction

Diets rich in fruit and vegetables have long been associated with reduced risk of chronic disease, particularly cardiovascular disease, cancers and type 2 diabetes [1]. Oxidative stress from increased amounts of reactive oxygen species (ROS) can cause extensive damage to cell structures, and is considered a major factor in the pathogenesis of these chronic diseases [2]. Evidence suggests that regular consumption of fruit and vegetables minimizes some of these harmful effects, which has been somewhat accredited to the presence of compounds possessing antioxidant properties [3]. The major antioxidants present in fruit and vegetables are: vitamin C, vitamin E, carotenoids and polyphenols, especially flavonoids, which all provide protection against free radicals [4]. The quality and quantity of these

antioxidant components are major attributes to the health benefits of fruit and vegetables that are associated with reduced risk of chronic disease [2]. For example, regular consumption of dark green leafy vegetables has shown a protective effect against two common eye diseases, cataract and macular degeneration, caused by free radicals generated by sunlight, metabolism and infection. These vegetables contain the pigments lutein and zeaxanthin, which accumulate in the eye and eradicate free radicals, thus preventing harm to the eye's sensitive tissues [5]. The high fibre content of fruit and vegetables also provide a protective effect. The bulking and softening action of indigestible fibre can reduce pressure inside the intestinal tract and calm the irritable bowel [6].

Broccoli (Brassica oleracea L. var. italica which originated from the eastern Plenck). Mediterranean region of Europe, is one of the most commonly consumed green vegetable worldwide. Broccoli possesses a wide range of bioactive compounds that have several health benefits, and are rich in both nutritional as well as non-nutritional antioxidants such as vitamins C and E, phenolic compounds, and glucosinolates (GSLs) {7-9]. Several epidemiological studies have shown that consumption of broccoli is positively associated with reduced risk of several types of cancers, type 2 diabetes, and cardiovascular diseases [10-11]. Furthermore, broccoli is known to possess antioxidant and anti-proliferative activities [3, 8]. These beneficial properties can be attributed to the presence of health-promoting phytochemicals such as GSLs, vitamins, carotenoids, phenols, flavonoids, and minerals [7, 9].

Phenolic compounds present in broccoli help neutralize or quench free radicals [12] and are often considered to be the most abundant antioxidants in the human diet [13]. Flavonoids and their derivatives possess antioxidant properties due to their ability to scavenge reactive oxygen species and inhibit oxidative stress [14]. All the antioxidants present in



broccoli show stronger interactive antioxidative properties when they work in groups as they function synergistically to reduce reactive oxygen species. The concentration of these phytochemicals and their properties in broccoli are dependent on both the genotype and environment. Several studies have significant confirmed changes in these phytochemicals based on plant genotype [9], growing season [15, 16], plant parts [17, 8], developmental stages [15], fertilization levels [18], temperature and irrigation [19], postharvest storage conditions [20], and even the cultivation year [21].

In addition to this, domestic cooking can dramatically reduce activities of antioxidant components, as many of these compounds are very sensitive to heat and are soluble in water [22]. Absorption of water during boiling can dilute and cause leaching of antioxidant compounds and thus decrease their antioxidant content [3]. There is a great amount of literature available concerning levels of antioxidant properties in raw fruit and vegetables [23-24, 2, 25]. However, there is less literature regarding the content of antioxidants in vegetables as usually eaten, i.e. after cooking. Variation in both cooking treatment and cooking duration may affect the nutritional value of vegetables [26]. Broccoli is normally cooked by boiling in water or microwaving before consumption [22]; thus, it is essential to determine which domestic cooking method and cooking duration are best for retaining antioxidants in this vegetable [27-28]. The aim of this study was to investigate the mineral content and to estimate the total flavonoid content of B. oleracea L.var. italica plenck.

1.1 Botanical Description

	•
Family :	Brassicaceae (Cruciferae)
Botanical name :	Brassica oleracea L.var.
	italica plenck.
English name :	broccoli
Myanmar name :	Pan Gaw Bi Sein
Part used :	Flower



Figure 1. Flower of *Brassica oleracea* L.var. italica plenck.

2. Materials and Methods

2.1 Collection and Preparation of Sample

The flowers of *B. oleracea* L.var. italica plenck. were collected from local Market, Chan Mya Thar Si Township, Mandalay Region, Myanmar. It was cut into small pieces and dried in the air for about four weeks. It was stored in a well-stoppered bottle and used for the determination of mineral content.

2.2 Determination of Mineral Content from the *Brassica oleracea* L.var. italica Plenck.

Mineral content of the *B. oleracea* L.var. italica plenck. was measured at the Department of Physic, University of Mandalay, Myanmar by applying EDXRF (Energy Dispersive X-ray Fluorescence Spectroscopy).

2.3 Extraction Procedure

100 g of fresh broccoli was crushed with 100 mL of 95 % EtOH by blender. These juices were squeezed, filtered and then centrifuged with 5000 rpm for 30 minutes. 105 mL of expressed juice which is the liquid product was obtained.

2.4 Qualitative Test for Flavonoids Ferric Chloride Test:

A few drops of neutral ferric chloride solution were added to 1 mL of extract solution. Formation of blackish red color indicates the presence of flavonoids.

Shinoda's Test:

To 1 mL of extract solution, a small piece of magnesium ribbon or magnesium foil was added and a few drops of concentrated HCl were added. Change in pink red colour shows the presence of flavonoids.

Lead- acetate Test:

To 1 mL of extract solution, a few drops of aqueous basic lead acetate solution were added. Formation of precipitate indicates the presence of flavonoids.

2.5 Quantitative Determination of Total Flavonoid Content

2.5.1 Principle

The basic principle of Aluminium chloride colorimetric method is that aluminium chloride forms acid stable complexes with the C-4 keto group and either the C-3 or C-5 hydroxyl group of flavones and flavonols. In addition it also forms acid labile complexes with the ortho-dihydroxyl groups in the A- or B- ring of flavonoids. Quercetin is reported to be suitable for building the calibration curve. Therefore standard quercetin solutions of various concentrations were used to build up the calibration curve [29-32].

2.5.2 Preparation and Determination of Standard Quercetin

10 mg of the standard quercetin was taken in a test tube. 100 mL of MeOH was added to the standard compound. The stock solution was obtained. It was diluted with MeOH in various ratios to obtained four ranges of concentration, such as 25 μ g/mL, 50 μ g/mL, 75 μ g/mL, and 100 μ g/mL respectively. Then, 4.0 mL of solution was prepared



_

for each concentration. 0.5 mL of each standard quercetin solution was taken in test tube and 1.5 mL methanol, 0.1 mL of 10% aluminium chloride, 0.1 mL of 1 M potassium acetate and 2.8 mL distilled water were added separately to each tubes.

These tubes were left at room temperature for 30 min after which the absorbance of the reaction mixture was measured at 415 nm with UV/ Visible spectrophotometer. The calibration curve was plotted by using the resulted absorbance data of standard quercetin solutions at concentrations 25 μ g/ mL to 100 μ g/ mL in methanol. The calibration curve of standard quercetin is shown in Figure 2 [29-32].

2.5.3 Determination of Total Flavonoid Content of *Brassica oleracea* L.var. italica Plenck.

The total flavonoid content of *B. oleracea* L.var. italica plenck. was measured by aluminium chloride (AlCl₃) according to the spectrophotometric method using quercetin as a standard. Firstly, 0.5 mL of extract solution was taken in test tube and 1.5 mL methanol, 0.1 mL of 10% aluminium chloride, 0.1 mL of 1M potassium acetate and 2.8 mL distilled water were added into tube.

This tube was left at room temperature for 30 min after which the absorbance of the reaction mixture was measured at 415 nm with UV/ Visible spectrophotometer. The assay was performed in triplicate. The total flavonoid content of *B. oleracea* L.var. italica plenck. was expressed as mg quercetin equivalent (QE) /100 g fresh weight [29-32].

3. Results and Discussion

3.1 Determination of Mineral Content from the Brassica oleracea L.var. italica Plenck.

The mineral contents of selected sample were described in Table 1.

Table	1.	Mineral	Contents	in	the	Brassicea	oleracea
L.var.italica Plenck.							

L.val.manca i lenck.					
No	Elements	Symbols	Result (%)		
1.	Potassium	K	2.73700		
2.	Phosphorus	Р	0.70070		
3.	Calcium	Ca	0.56290		
4.	Sulphur	S	0.37730		
5.	Chlorine	Cl	0.33400		
6.	Aluminium	Al	0.18420		
7.	Iron	Fe	0.02119		
8.	Silicon	Si	0.02100		
9.	Zinc	Zn	0.00743		
10.	Titanium	Ti	0.00691		
11.	Manganese	Mn	0.00555		
12.	Rubidium	Rb	0.00534		
13.	Barium	Ba	0.00340		
14.	Vanadium	V	0.00282		
15	Conner	Cu	0.00108		

According to the resulted data, broccoli consists of K, P, Ca, S, Cl, Al, Fe, Si, Zn, Ti, Mn, Rb, Ba, V, and Cu, respectively. Among them, it was found that potassium (K), Phosphorus (P), calcium (Ca), sulphur (S) and chlorine (Cl) contain the higher amount than other in this sample and moderate contents of aluminium (Al), iron (Fe) and silicon (Si) could be observed.

3.2 Estimation of Total Flavonoid Content in *Brassica oleracea* L.var. italica Plenck. Special Test for Flavonoid

The fresh juice obtained by crushing the *B. oleracea* L.var. italica plenck. was examined by using the special qualitative tests of flavonoid. The resulted data were tabulated in Table 2.

Table 2. The results of qualitative test for flavonoid							

No	Experiment	Observation	Inference
1.	Ferric Chloride Test:	Blackish red colour was appeared	Flavonoid may be present
2.	Shinoda's Test:	Colour turns to pink red colour	Flavonoid is present
3.	Lead Acetate Test:	Precipitate was produced	Flavonoid is present

From these results, it was observed that the selected sample consists of flavonoid compounds.

The calibration curve was plotted against by using the resulting data of standard quercetin solution as shown in Figure 2.



Figure 2: Concentration absorbance calibration curve for standard quercetin

In addition, the total flavonoid content of the *B. oleracea* L.var. italica plenck. was carried out by aluminium chloride spectrophotometric method using the quercetin as a standard. The absorbance values of prepared sample solutions were measured by UV spectrophotometer at 415 nm with respect to the blank solution. From these results, the amount of total flavonoid content of analyzed sample was obtained by using the standard graph. The results are described in Table 3.



solutions of <i>B. oleracea</i> L.var. italica Plenck.					
					Flavonoid

Table 3 The results of total flavonoid content of extract

No	Name of Sample	Flavonoid (mg/100 g)	(mg/100 g) Mean ± Standard Deviation	
	B. oleracea	10.17		
1.	L.var. italica Plenck.	10.34	10.17 ± 0.16	
		10.01		

The total flavonoid content present in the selected vegetable (Broccoli) was found as 10.17 ± 0.16 mg quercetin equivalent (QE) per 100 g fresh weight.

4. Conclusion

In this research work, one of Myanmar well known vegetables, B. oleracea L. var. italica plenck., Myanmar name Pan Gaw Bi Sein, Broccoli, which is flavonoid rich vegetable, was selected to determine the mineral content and to evaluate the total flavonoid present in it qualitatively and quantitatively. The mineral content of selected sample was determined by EDXRF method. It was found that potassium contains the highest amount in this sample. In accordance with the qualitative test of flavonoid, it was confirmed that this fresh juice contains the flavonoid compounds. Moreover, the total flavonoid content of the extract obtained from the selected sample could be evaluated by UV spectrophotometer using the aluminium chloride method at 415 nm. It was found that the total flavonoid content of B. oleracea L. var. italica plenck. is 10.17 ± 0.16 mg quercetin equivalent (QE) per 100 g fresh weight. The resulted data of the current study showed that the selected sample, the broccoli, had the considerable amount of total flavonoid compounds. Flavonoid compounds that are secondary metabolites are antioxidants. Quercetin, the most abundant dietary flavanol, is a potent antioxidant because it has all the right structural feactures for free radical scavening activity.

5. Acknowledgements

We are deeply thankful to Dr Kyae Mon Lwin, Professor, Head of Department of Chemistry, Kyaukse University, Mandalay Region, Myanmar for her kind permission and for providing research facilities.

References

- [1] Faller, A. L. K. and Fialho, E. The antioxidant capacity and polyphenol content of organic and conventional retail vegetables after domestic cooking, *Food Research International*, 2009, 42, 210–215.
- [2] Roy, M. K., Juneja, L. R., Isobe, S. et al. Steam processed broccoli (Brassica oleracea) has higher

antioxidant activity in chemical and cellular assay systems, *Food Chemistry*, 2009, 114 (1), 263–269.

- [3] Podsedek, A. Natural antioxidants and antioxidant capacity of Brassica vegetables: a review, *Swiss Society* of Food Science and Technology, 2007, 40, 1–11.
- [4] Monero, D. A., Perez-Balibrea, S., Ferreres, F. et al. Acylated anthocyanins in broccoli sprouts, *Food Chemistry*, 2010, 123 (2), 358–363.
- [5] Christen, W. G., Lui, S., Glynn, R. J., *et al.* Dietary carotenoids, vitamins C and E, and risk of cataract in women, *Arch Ophthalmol*, 2008, 126, 102–109.
- [6] Lembo, A. and Camilleri, M. Chronic constipation, New England *Journal of Medicine*, 2003, 349, 1360– 1368.
- [7] Aires, A., Fernandes, C., Carvalho, R., Bennett, R.N., Saavedra, M.J., and Rosa, E.A.S. Seasonal effects on bioactive compounds and antioxidant capacity of six economically important *Brassica* vegetables. Molecules, 2011, 16:6816-6832.
- [8] Bhandari, S.R., and Kwak, J.H. Chemical composition and antioxidant activity in different tissues of *Brassica* vegetables. Molecules, 2015, 20:1228-1243.
- [9] Jo, J.S., Bhandari, S.R., Kang, G.H., and Lee, J.G. Comparative analysis of individual glucosinolates, phytochemicals, and antioxidant activities in broccoli breeding lines. Horticulture Environment and Biotechnology, 2016, 57(4):392-403.
- [10] Razis, A.F.A., and Noor, N.M. Cruciferous vegetables: dietary phytochemicals for cancer prevention. Asian Pacific Journal of Cancer Prevention, 2013, 14(3):1565-1570.
- [11] Bachiega, P., Salgado, J.M., De Carvalho, J.E., Ruiz, A.L.T.G., Schwarz, K., Tezotto, T., et al. Antioxidant and antiproliferative activities in different maturation stages of broccoli (*Brassica oleracea* Italica) biofortified with selenium. Food Chemistry, 2016, 190:771-776.
- [12] Cartea, M.E., Francisco, M., Soengas, P., and Velasco, P. Phenolic compounds in *Brassica* vegetables. Molecules, 2011, 16:251-280.
- [13] Faller, A.L.K., and Fialho, E. The antioxidant capacity and polyphenol content of organic and conventional retail vegetables after domestic cooking. Food Research International, 2009, 42:210-215.
- [14] Pourcel, L., Routaboul, J.M., Cheynier, V., Lepiniec, L., and Debeaujon, I. Flavonoid oxidation in plants: from biochemical properties to physiological functions. Trends in Plant Science, 2007, 12(1):29-36.
- [15] Vallejo, F., Garcia-Viguera, C., and Tomas-Barberan, F. Changes in broccoli (*Brassica oleracea* L. var. *italica*) healthpromoting compounds with inflorescence development. Journal of Agricultural and Food Chemistry, 2003, 51:3776-3782.
- [16] Bhandari, S.R., and Kwak, J.H. Seasonal variation in phytochemicals and antioxidant activities in different tissues of various *Broccoli* cultivars. African Journal of Biotechnology, 2014, 13(4):604-615.
- [17] Perez-Balibrea, S., Moreno, D.A., and Garcia-Viguera, C. Genotypic effects on the phytochemical quality of seeds and sprouts from commercial broccoli cultivars. Food Chemistry, 2011, 125(2):348-354.
- [18] Fabek, S., Toth, N., Redovnikovic, I.R., Custic, M.H., Benko, B., and Zutic, I. The effect of nitrogen fertilization on nitrate accumulation, and the content of minerals and glucosinolates in broccoli cultivars.



Food Technology and Biotechnology, 2012, 50(2):183-191.

- [19] Pek, Z., Daood, H., Nagyne, M.G., Berki, M., Tothne, M.M., Nemenyi, A., et al. Yield and phytochemical compounds of broccoli as affected by temperature, irrigation, and foliar sulfur supplementation. HortScience, 2012, 47(11):1646-1652.
- [20] Fernandez-Leon, M.F., Fernandez-Leon, A.M., Lozano, M., Ayuso, M.C., and Gonzalez-Gomez, D. Different postharvest strategies to preserve broccoli quality during storage and shelf life: controlled atmosphere and 1-MCP. Food Chemistry, 2013, 138(1):564-573.
- [21] Jo, J.S., Bhandari, S.R., and Lee, J.G. Yearly variation in glucosinolate content in inflorescence of broccoli breeding lines. Horticultural Science and Technology, 2018, 36(3):406-416.
- [22] Zhang, D. and Hamauzu, Y. Phenolics, ascorbic acid, carotenoids and antioxidant activity of broccoli and their changes during conventional and microwave cooking, *Food Chemistry*, 2004, 88 (4), 503–509.
- [23] Sun, T., Powers, J. R. and Tang, J. Evaluation of the antioxidant activity of asparagus, broccoli and their juices, *Food Chemistry*, 2007, 105 (1), 393–401.
- [24] Gawlik-Dziki, U. Effect of hydrothermal treatment on the antioxidant properties of broccoli (*Brassica* oleracea var. botrytis italica) florets, Food Chemistry, 2008, 109 (2), 393–401.
- [25] Lemoine, M. L., Chaves, A. R. and Martinez, G. A. Influence of combined hot air and UV-C treatment on the antioxidant system of minimally processed broccoli (*Brassica oleracea* L. var Italica), *Food Science and Technology*, 2010, 43 (9), 1313–1319.
- [26] Lin, C. H. and Chang, C. Y. Textural change and antioxidant properties of broccoli under different

cooking treatments, *Food Chemistry*, 2005, 90 (1–2), 9–15.

- [27] Gliszczynska-Swiglo, A., Ciska, E., Pawlak-Lemanska, K. *et al.* Changes in the content of healthpromoting compounds and antioxidant activity of broccoli after domestic processing, *Food Additives and Contaminants*, 2006, 23 (11), 1088–1098.
- [28] Gebczynski, P. and Lisiewsk, A. Comparison of the level of selected antioxidant compounds in frozen broccoli produced using traditional and modified methods, *Innovative Food Science and Emerging Technologies*, 2006, 7, 239–245.
- [29] Akbay P, Basaran AA, Undeger U, Basaran N. In vitro immune modulatory activity of flavonoid glycosides from Utrica dioica L, Phytother Res., 2003, 17 : pp. 34-37.
- [30] Kaufman PB, Cseke LJ, Warber S, Duke JA, Brielmann. Natural products from plants, CRC press, New York, 1999, pp. 20-22.
- [31] Arnt Win, Aye Mon Thida Nyo. Evaluation of Total Phenolic and Flavonoid Content of Pomegranate Juice. Journal of Emerging Technologies and Innovative Research (JETIR). 2019, 6: pp. 431-436.
- [32] G.C. Bag, P. Grihanjali Devi & Th. Bhaigyabati, Assessment of Total Flavonoid Content and Antioxidant Activity of Methanolic Rhizome Extract of Three Hedychium Species of Manipur Valley. Int. J. Pharm. Sci. Rev. Res., 2015, 30(1) : pp. 154-159