Photoproduction of Hydrogen by Vishakapatnam Photosynthetic Bacteria and their Effect on Different Cultural Conditions

Jahnavi Alwala¹, M.P.Pratap Rudra^{*1}, Ramchander Merugu²

¹Department of Biochemistry, Osmania University, Hyderabad ²University College of Science, Mahatma Gandhi University, Nalgonda Corresponding Author*:*jahnavichandu.enjapuri@gmail.com*

Abstract:

Hydrogen, well advised by many scientists, as an ecofriendly and optimal fuel for the imminent future, were well oust plentiful applications. in Rhodopseudomonas species have shown an immense grade of morphological and structural analogy, however, they are phylogenetically quite diverse. Therefore, the present study for Hydrogen production affirms that the influence consortium of purple non sulphur bacteria isolated from Vishakapatnam marine water. The different parameters like various Carbon, Nitrogen, Growth factors and P_H variation was identified for hydrogen production by the anoxygenic bacteria. The amount of hydrogen produced varies with various carbon, nitrogen, growth factors and P_H used in the medium. The bacterial culture was cultured in 50ml vessel within 30ml of culture to produce hydrogen in anoxygenic conditions and incubated in 2000lux light intensity for 196 hours. The best result for highest hydrogen productivity was 6ml/30ml of sodium benzoate, ammonium chloride, cyanocobalamine culture and at P_H 7.0, 7.5 to purple non sulphur bacteria isolated from Vishakapatnam marine water.

Keywords:

Purple non sulphur bacteria; Hydrogen production; carbon; nitrogen sources; growth factors; P_H

Introduction:

In consonance with sustainable expansion and waste minimization issues, biological hydrogen production from renewable sources, also known as "green technology" has revived attention in

Morphological, recent years. cytological and physiological properties of the genus Rhodopseudomonas have been outlined [1-3].Marine habitats serve fine niches for anoxygenic phototrophic bacteria, which are extensively assigned in diverse coastal marine habitats. Most common anoxygenic phototrophic bacteria have been isolated from estuarine salt pans, salt marshes, coastal lagoons with elevated salt concentrations, tidal waters; brackish waters and marine coastal sediments .They have even been found in the extreme marine habitats of Antarctica [4-11]. The peculiar substrates such as acetate, lactate, benzoate, malate, mannitol and starch are outlined by phototrophic bacteria as an electron donor for hydrogen production. Impact of various cultural conditions on hydrogen production is expressed[12-15].Specially from hydrogen production multifold by-products are produced from photosynthetic bacteria makes a continuity for our earlier work on biotechnological applications[16-26].The present aspects of the above data, reveals the impact of cultural conditions like various carbon, nitrogen, growth factors and pH variation on hydrogen production by anoxygenic bacterial consortium isolated from Vishakapatnam marine water was examined and explained.

Materials and Methods:

Photoproduction of Hydrogen by Vishakapatnam Photosynthetic Bacteria and their Effect on Different Cultural Conditions Jahnavi Alwala1, M.P.Pratap Rudra, Ramchander Merugu Page | 805



Purple non Sulphur an oxygenic Phototrophic bacterium was isolated from marine water samples by enrichment techniques by inoculating into the medium and incubated an aerobically under 2000 lux light intensity. A selected consortium isolate from Vizag marine water that had both screening tests passed was characterized using both morphological and physiological properties and identified according Bergey's to Manual of Systematic Bacteriology (1994).Growth conditions were measured at 660nm Optical density under **UV-VIS** Spectrophotometer. The harvested cells were obtained from the 30ml of bacterial culture centrifuged (10,000xg for 30mins) and washed with 0.3% saline and cells were suspended in basal medium. Based on experimental conditions provided at different concentrations of electron donors, nitrogen sources, ten days old cultures of phototrophic bacteria of 1 % (v/v)concentration into the basal medium were inoculated containing carbon sources Sodium

benzoate,Glucose,Galactose,Mannose,Ara binose,Lactose,Mannitol,Malic acid, Citric acid, Sodium succinate with ammonium chloride as nitrogen source. Ammonium chloride, ammonium nitrate,glycine,sodium

glutamate, histidine, tryptophan, tyrosine, thr eonine, alanine with sodium benzoate as carbon source, different growth factors like panthothenic acid, nicotinic acid, biotin, folic

acid,riboflavin,cyanocobalamine and basal medium at different PH variations from 6.5,7,7.5,8,8.5,9,9.5 were also explored. The incubation period was 196 hours after inoculation of the consortium. The technique used for hydrogen measurement was water displacement method where as Gas Chromatography was used for gas analysis.

Results and Discussion:

Ten day active cultures were used to assess their probability of producing hydrogen was shown in Table 1 and Figure **1**.Photosynthetic bacterial consortium produced varying amounts of hydrogen with various carbon, nitrogen, growth factors and variation in P_H under anaerobic light. Sodium benzoate, mannose, lactose and mannitol showing maximum and followed by equal volumes in glucose,galactose,arabinose and malic acid were good carbon sources for production of hydrogen by photosynthetic consortium. bacterial Maximum production of hydrogen was 6ml /30ml culture was produced in presence of sodium benzoate. Effect of various nitrogen sources on hydrogen production was shown in Table 2 and Figure 2.In the of anaerobic conditions, presence ammonium chloride produces 6ml/30ml culture which shows a maximum amount of hydrogen followed by ammonium nitrate, sodium glutamate and tryptophan. Thus, maximum hydrogen production of both the carbon and nitrogen sources was observed in sodium benzoate, ammonium chloride. Glucose and galctose shows equal volumes of hydrogen and ammonium nitrate, sodium glutamamte, tryptophan has shown the equal volumes of hydrogen production. Impact of P_H variation on hydrogen production was Table 3 described in and Figure 3.Maximum amount of hydrogen production was observed in 7.0 and 7.5 P_{H} Biotin, folic acid and riboflavin showed the equal amounts and maximum hydrogen production was observed in nicotinic acid and cyanocobalamine was shown in Table 4 and Figure 4.



Table 1. Effect of various Carbon Sources on Hydrogen Production (ml/30ml culture)

Optical Density (in OD)	Carbon /Electron Donor	Hydrogen (in ml)
1.0655	Sodium Benzoate	6±0.0
0.5179	Glucose	5.5±0.1
0.7593	Galactose	5.5 ± 0.2
0.8357	Mannose	6.0±0.1
0.7806	Arabinose	5.5±0.2
0.7959	Lactose	6±0.1
0.7226	Mannitol	6±0.0
0.9322	Malic acid	5.5±0.1
0.2498	Citric acid	4.0±0.6
0.3587	Sodium succinate	4.5±0.4



Table 2: Effect of various Nitrogen sources on Hydrogen production (ml/30ml)

Nitrogen Source	Optical Density (in OD)	Hydrogen (in ml)
Ammonium Chloride	1.9049	6±0.2
Ammonium Nitrate	0.8130	5.5 ± 0.2
Glycine	0.8690	6.0±0.2
Sodium glutamate	0.5065	5.5 ± 0.4
Histidine	0.3548	4 ± 0.5
Tryptophan	0.4546	4.5±0.2
Tyrosine	0.8495	6±0.0
Threonine	0.9924	6±0.1
Alanine	0.7979	6±0.1



Table 3: Effect of various P_H on Hydrogen Production (ml/30ml)

P _H	Optical Density (in OD)	Hydrogen (in ml)		
P _H 6.5	1.0709	5.5±0.2		
P _H 7.0	1.5063	6.0±0.0		
P _H 7.5	1.7990	6.0±0.1		
P _H 8.0	1.1943	5.0±0.1		
P _H 8.5	0.8727	4.5±0.4		
P _H 9.0	1.4099	5.5±0.1		
P _H 9.5	1.1444	5.0±0.3		



Photoproduction of Hydrogen by Vishakapatnam Photosynthetic Bacteria and their Effect on Different Cultural Conditions Jahnavi Alwala1, M.P.Pratap Rudra, Ramchander Merugu Page | 808



International Journal of Research (IJR) Vol-1, Issue-8, September 2014 ISSN 2348-6848

Table 4: Effect of various Growin Factors on Hydrogen Droduction (mi/jum	Table	4: Effect	t of various	Growth	Factors on	Hvdrogen	production ((ml/30ml
--	-------	-----------	--------------	--------	------------	----------	--------------	----------

Growth Factors	Optical Density (in OD)	Hydrogen (in ml)
Panthothenic acid	0.8116	5.0±0.5
Nicotinic acid	1.6458	6.0±0.1
Biotin	1.0011	5.5±0.3
Folic acid	1.0394	5.5±0.3
Riboflavin	1.0051	5.5±0.2
Cyanocobalamine B ₁₂	1.1737	6.0±0.1



Conclusion:

The present study reveals that impact of various carbon, nitrogen, P_H variation and growth factors shown that equals and maximum amounts of hydrogen production was shown in sodium benzoate, ammonium chloride, P_H 7.0, 7.5 and nicotinic acid, cyanocobalamine and less amounts of hydrogen production in citric acid, histidine and panthothenic acid and at P_H 8.5.

Acknowledgement:

The authors are gratefully acknowledged to the University Grants Commision,New Delhi for their financial assistance and Department of Biochemistry,Osmania University. **References:**

[1] Imhoff JF, Tru["] per HG, Pfennig N. Int J Bacteriol 1984; 34(3):340–3.

[2] Oda, Y., Star, B., Huisman, L.A., Gottschal, J.C., and Forney, L.J., 2003. Biogeography of the purple nonsulfur bacterium *Rhodopseudomonas palustris*. *Appl. Environ. Microbiol.*, Vol. 69, No. 9, pp. 5186-5191.

[3] Zhou, X.X, Wang, Y.B., Li, W.F., and Pan. Y.J. 2007. In vitro assessment of gastrointestinal viability of two photosynthetic bacteria, *Rhodopseudomonas palustris* and *Rhodobacter sphaeroides*. J. Zhejiang Univ. SCIENCE B., Vol. 8, No. 9, pp. 677-683.

[4] Imhoff, J. F. (1983). Rhodopseudomonas marina sp. nov., a new marine phototrophic purple bacterium. Syst Appl Microbiol 4, 512–521.

Photoproduction of Hydrogen by Vishakapatnam Photosynthetic Bacteria and their Effect on Different Cultural Conditions Jahnavi Alwala1, M.P.Pratap Rudra, Ramchander Merugu P a g e | 809

International Journal of Research (IJR) Vol-1, Issue-8, September 2014 ISSN 2348-6848

[5] Imhoff, J. F. (1988). Anoxygenic phototrophic bacteria. In Methods in Aquatic Bacteriology, pp. 207–240. Edited by B. Austin. Chichester, New York: Wiley.

[6] Imhoff, J. F. (2001). Transfer of Rhodopseudomonas acidophila to the new genus Rhodoblastus as Rhodoblastus acidophilus gen. nov., comb. nov. Int J Syst Bacteriol Microbiol 51, 1863–1866.

[7] Imhoff, J. F. & Pfennig, N. (2001). Thioflavicoccus mobilis gen. nov., sp. nov., a novel purple sulfur bacterium with bacteriochlorophyll b. Int J Syst Bacteriol Microbiol 51, 105–110.

[8] Imhoff, J. F., Su⁻ ling, J. & Petri, R. (1998a). Phylogenetic relationships among the Chromatiaceae, their taxonomic reclassification and description of the new genera Allochromatium, Halochromatium, Isochromatium, Marichromatium, Thiococcus, Thiohalocapsa and Thermochromatium. Int J Syst Bacteriol 48, 1129–1143.

[9] Imhoff, J. F., Su[¬] ling, J. & Petri, R. (1998b). Reclassification of species of the spiralshaped phototrophic purple non-sulfur bacteria of the a-Proteobacteria : description of the new genera Phaeospirillum gen. nov., Rhodovibrio gen. nov., Rhodothalassium gen. nov. and Roseospira gen. nov. as well as transfer of Rhodospirillum fulvum to Phaeospirillum fulvum comb. nov., of Rhodospirillum molischianum to Phaeospirillum molischianum comb. nov., of Rhodospirillum salinarum to Rhodovibrio salinarum comb. nov., of Rhodospirillum sodomense to Rhodovibrio sodomensis comb. nov., of Rhodospirillum salexigens to Rhodothalassium salexigens comb. nov. and of Rhodospirillum mediosalinum to Roseospira mediosalina comb. nov. Int J Syst Bacteriol 48, 793–798.

[10] Madigan, M. T., Jung, D. O., Woese, C. R. & Achenbach, L. A. (2000). Rhodoferax antarcticus sp. nov., a moderately psychrophilic purple non-sulfur bacterium isolated from an Antarctic microbial mat. Arch Microbiol 173, 269–277.

[11] Karr, E. A., Sattley, W. M., Jung, D. O., Madigan, M. T. & Achenbach, L. A. (2003). Remarkable diversity of phototrophic purple bacteria in a permanently frozen Antarctic lake. Appl Environ Microbiol 69, $4910-4914.\pm$

[12] H. Koku, I. Eroglu, U. Gndz, M. Ycel, L. Trker, International Journal of Hydrogen Energy 28 (2003) 381 – 388.

[13] G. Zheng, L. Wang, Z. Kang, Renewable Energy 35 (2010) 2910 – 2913.

[14] F. R. Hawkes, R. Dinsdale, D. L. Hawkes, I. Hussy, International Journal of Hydrogen Energy 27 (2002) 1339 – 1347.

[15] M. R. Melnicki, E. Eroglu, A. Melis, International Journal of Hydrogen Energy 34 (2009) 6157 – 6170.

[16] Ramchander, M., Pratap, M. P., Nageshwari, B., Girisham, S., and Reddy, S. M. (2012). Factors influencing the production of hydrogen by the hydrogen by the purple non-sulphur phototrophic bacterium *Rhodopseudomonas acidophila* KU001. *Microb. Biotechnol.*, **5**: 674-678.

[17] Ramchander Merugu, Vasantha Mittapelli, M.P. Pratap Rudra, S. Girisham, S.M. Reddy (2012). Photoproduction of Hydrogen by Alginate Immobilized *Rhodobacter capsulatus* KU002 under Sulphate and Phosphate Limitations. International Journal of Environment and Bioenergy, 4(3): 141-146.

[18] Ramchander Merugu, Vasantha Mittapelli, Pratap Rudra Manthur Padigya, Girisham Sivadevuni, Reddy Solipuram Madhusudhan (2013). Photoproduction of Hydrogen by Alginate Immobilised Cultures of *Rhodobacter capsulatus* KU002 Isolated from Tannery Effluents. Journal of Biofuels. 4 (2): 56-60.

[19] Ramchander Merugu, M. P. PratapRudra, B.Nageshwari, A. Sridhar Rao, and D.Ramesh. Photoproduction of Hydrogen under Different Cultural Conditions by Alginate Immobilized *Rhodopsedomonas palustris* KU003. ISRN Renewable Energy. Volume 2012, Article ID 757503, 5 pages doi:10.5402/2012/757503.



[20] Ramchander Merugu, M.P.Pratap Rudra, S.Girisham and S.M.Reddy (2013). Bioproduction of hydrogen by alginate immobilized *Rhodopsedomonas acidophila* KU001. International Journal of Chemical Engineering and Applied Sciences 3(1): 7-9.

[21] Ramchander, M., M.S.K.Prasad, Vasavi, DS. Girisham and S.M. Reddy (2007). Bioremediation of waste water by two Anoxygenic Phototrophic bacteria *Nat.Acad. Sci.* Lett.30:223-227.

[22] Ramchander Merugu, M.S.K. Prasad, S. Girisham and S.M.Reddy (2008). Effect of trace elements on the growth of two Anoxygenic phototrophic bacteria *Ecol. Envi. Con.* 14; 367-369.

[23] Ramchander Merugu, M. P. Pratap Rudra, A. Sridhar Rao, D. Ramesh, B. Nageshwari, K. Rajyalaxmi, S. Girisham, and S. M. Reddy (2011). Influence of Different Cultural Conditions on Photoproduction of Hydrogen by *Rhodopseudomonaspalustris* KU003. *ISRN Renewable Energy*, 328984-90.

[24] Ramchander Merugu, M.P.Pratap Rudra, Atthapu Thirupathaiah and Veerababu Nageti (2011). Hypocholesterolemic effect of the anoxygenic phototrophic bacterium *Rhopseudomonas palustris* MGU001 in hen laying eggs. International Journal of Applied Biology and Pharmaceutical Technology. 2 (2): 463 to 466.

[25] Ramchander Merugu, M.P.Pratap Rudra, Atthapu Thirupathaiah, S.Girisham and S.M.Reddy (2011). Optimisation of Indole Acetic Acid Production by two Anoxygenic Phototrophic bacteria Isolated from Tannery effluents. Journal of pure and applied Microbiology. 5(2): 34-37.

[26] Ramchander Merugu, M.P.Pratap Rudra, Atthapu Thirupathaiah, S.Girisham and S.M.Reddy (2011). Chromate Reduction by a Purple non Sulphur Phototrophic Bacterium *Rhodobacter capsulatus* KU002 Isolated from Tannery Effluents. Journal of Pure and Applied Microbiology. 5(2): 66-69.