

Fecundity Of *Synodontis Membranaceus* (Geoffroy Saint-Hilaire, 1809) In The Lower River Benue At Makurdi.

Akombo, P. M.,¹cheikyula, J. O.,² Atile, J. I.¹

¹Department of Biological Sciences, Benue State University, PMB 102119 Makurdi, Nigeria.

²Department of Fisheries and Aquaculture, Federal University of Agriculture Makurdi, Nigeria
mbakaanpauline@yahoo.com;

ABSTRACT

An investigation was carried out on the fecundity and Gonado-somatic index (GSI) of *Synodontis membranaceus* from River Benue over a 24 month period from January, 2009 to December, 2010. A total of 106 fish were collected: 59 (55.66%) males; 47 (44.24%) females. 36 of the females had developing eggs of stages IV-VI. The standard lengths of the 36 females ranged from 18.40cm - 28.10cm with the mean of 23.34±0.39. Total weight ranged from 164.10g - 556.70g mean of 314.20±16.00; gonad weight ranged from 3.17g to 90.00g mean of 21.76±3.01. GSI ranged from 1.02 - 17.53 mean of 6.73±0.68 and absolute fecundity ranged from 1,327 to 119,954 mean of 33,173±15.36. Fecundity was found to correlate positively with gonad weight ($r^2=0.7535$) but not with GSI ($r^2=0.5728$) and body weight ($r^2=0.2833$).

Key words: *Synodontis membranaceus*, Fecundity, Gonado-somatic (GSI) index, Lower River Benue.

INTRODUCTION

Synodontis membranaceus is one of the species of the genus *Synodontis*. Teugels (1996) reported that this species was endemic to Africa. It is one of the most important commercial fish species in the artisanal fisheries in Nigeria (Reed *et al.*, 1967). In River Benue it is not common throughout the year but it is one of the species that attains the biggest size. Because of their relatively large sizes they are generally preferred by fishermen and consumers and command higher market prices. *S. membranaceus* has a potential for culture (Owolabi, 2005). They are popular aquarium species (Teugels, 1996; Owolabi, 2008) as they exhibit the habit of swimming upside down. Bishai and Abu Gideiri (1965b) carried out on feeding and reproduction on *S. membranaceus* in River Nile at Khartoum; Imevbore (1970) on sex ratio and fecundity in River Niger, Nigeria prior to impoundment; Other studies were on orientation and Swimming by Willoughby (1974), and Owolabi (2005) on other aspects of the biology of the fish in Jebba Lake, Nigeria. In the lower River Benue, Akombo *et al.*, (2011) worked on its Morphometric and growth pattern. This work is aimed at determining the

fecundity of *S. membranaceus* in River Benue at Makurdi.

Fecundity is the number of eggs produced by a fish (Lagler *et al.*, 1977). Absolute fecundity is the number of ripe eggs found in the female just prior to the next spawning period. (Bagenal, 1978); Halim and Guma'a, 1989). The number of eggs developing in a female just prior to spawning and the survival of early stages after hatching plays an important part in fish production, population dynamics, mortality, and survival rates. For the evaluation of the commercial potential of the stock, life history, culture and management of a fishery, the knowledge about fecundity of a fish is essential.

MATERIALS AND METHODS

Study Area

The study was carried out in the Lower Benue River at Makurdi, Nigeria. The Lower Benue River is the portion of the Benue River that is contained within the Benue and Kogi States of Nigeria (Reid and Sydenhan, 1979). River Benue originates from the Adamawa Mountains of Cameroun and flows west across East-Central Nigeria (Nedeco, 1959). It joins at the River Niger Lokoja Nigeria which flows to the Atlantic Ocean.

The River has extensive alluvial plain stretching for many kilometers, which covers a distance of approximately 187 kilometers. The extensive flood plain forms breeding grounds for many fish species (Beadle, 1974). The highest water levels are in August to September and the Lowest are in March to April.

Sample Collection

S. membranaceus were purchased from the fish sellers at Wadata Market, Makurdi, which is one of the fish landing sites on the bank of River Benue. The fish were caught by fishermen on the river using different types of gear such as gill nets, cast nets, hooks and traps. The fishes were procured fortnightly for 24 months (Jan. 2009 to Dec. 2010) and transported to the Biology Laboratory in Benue State University for identification and measurements. Identification was done using the keys of Reed *et al.*, (1967) Holden and Reed (1972), Babatunde and Raji (1998) and Idodo-Umeh (2003).

Length and Weight Measurements:

The standard lengths (SL) of the fish were measured using a measuring board. The anterior tip of the fish was

placed against a stop at the beginning of the measuring scale with its mouth closed. SL was the length from the tip of the snout to the base of the tail fin rays and this was measured to the nearest 0.1 centimeter. The total weight (TWT) was measured using a digital electronic weighing balance (Adam AFP 4100L). This was read to the nearest 0.1 gramme.

Sex Determination

The sex of *Synodontis* species were identified from internal sex organs after dissection. The gonads were inspected and Examined using the keys of Nikolsky (1963). The maturing stages of the gonads were classified according to the classification of Kesteven (1960).The stages were : **Stage I**, immature; **Stage II**, developingeggs; **Stage III**, still developingeggs; **Stages IV-V** mature eggs; **VI** ripe-running eggs and **Stage VII**, spent fish. The female gonads in gonad stages of **IV-VI** were removed and preserved in Gilson's fluid as described in Bagenal and Tesch(1978).

FECUNDITY STUDIES

The fecundity of the fish was determined using the female gonads in developmental stages of IV-VI. The ovaries were removed, and preserved in labelled reagent bottles containing Gilson's fluid at room temperature for 12 weeks to liberate and harden the eggs. The Gilson's fluid was prepared by adding 100 ml of 60% alcohol, 15ml of 80% nitric acid, 18 ml of glacial acetic acid, 20g of mercuric chloride and made to 1000ml with distilled water (Bagenaland Tesch, 1978).

While in storage, they were vigorously shaken at intervals until all the eggs were released from the ovarian tissues. After storage the Gilson's fluid decanted. The ovarian tissues were removed by successive shaking of the eggs and decanting the water. The clean

eggs were poured on filter paper to pick the remaining ovarian tissues and drain the water.

FECUNDITY ESTIMATION

The eggs were dried in an oven at 40⁰ C for 24 hours. Counting was done manually for five similar subsamples. The mean number of eggs in the five subsamples gave the number of eggs per 0.1gram weight. Fecundity was calculated by multiplying the total weight of eggs by the number of eggs per gram weight to give the absolute fecundity:

Fecundity, F=Total Weight of eggs × 750

The size of the eggs was determined by measuring the diameter of 100 eggs, randomly selected per female fish. Measurement of egg diameter was done with a calibrated micrometer mounted in the eyepiece of a binocular microscope Lagler (1971).

GONADO-SOMATIC INDEX

The gonado-somatic index (GSI) was calculated based on the formula by Lagler (1971):

$$GSI = \frac{\text{Gonad weight} \times 100}{\text{Total body weight}}$$

RESULTS

A total number of 106 specimens of *S.membranaceus* were used for this study, out of which 59 (55.66%) were males while 47 (44.34%) were females. Out of the 47(44.34%) females, 36 (76.60) had developing eggs of stages IV-VI (plate 1- 8) which were used for the fecundity estimation.



Plate 1: STAGE I- IMMATURE GONADS 2: STAGE II - DEVELOPING EGGS



Plate 3:

STAGE III -DEVELOPING EGGS Plate 4: STAGE IV - MATURING EGGS



Plate 5: STAGE V - MATURING EGGS Plate 6: STAGE VI-RIPE-RUNNING EGGS



Plate 7: STAGE VII: SPENT

Plate 8: MALE GONADS

The number of eggs in the female ranging from 18.40cm to 23.30cm weighing 164.10g- 314.10g was

1,327eggs and the female ranging from 27.50cm- 28.10cm with the weight of 314.30g to 556.69g was

119,954 eggs with the mean of $33,173 \pm 15.36$ (Table 1). These observations indicate that *S.membranaceus* is a species that is highly fecund. The low fecundity

observed in fish of 18.40cm to 23.30cm could have been that the specimen had already shaded its eggs.

Table 1: Morphometric and Reproductive Parameters of *S.membranaceus* in Lower River Benue

Parameter	Min	Max	Mean±SE
Standard Length (cm)	18.40	28.10	23.34±0.39
Total Weight (g)	164.10	556.70	314.20±16.00
Gonad Weight (g)	3.71	90.00	21.76±3.01
Gonadosomatic Index (GSI)	1.02	17.53	6.73±0.68
Fecundity	1,327	119,954	33,173±15.36

Regression graphs of standard length-body weight (Fig 1), fecundity-body weight (Fig 2) fecundity-gonad weight (Fig 3) and fecundity-GSI relationships shows that there was positive correlation ($P < 0.01$) between Standard length and body weight ($r^2 = 0.7838$), Fecundity

and gonad weight ($r^2 = 0.7535$) but weak positive correlation in GSI and fecundity ($r^2 = 0.5727$). Fecundity and body weight were not correlated ($r^2 = 0.2833$).

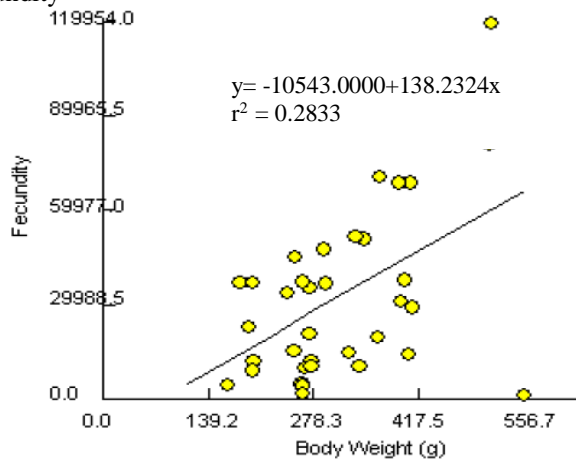
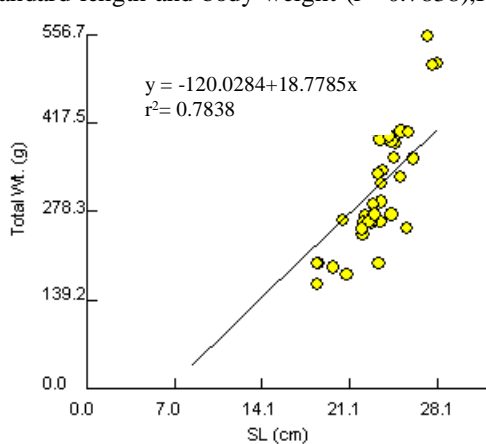


Fig.1: Body weight VS SL Fig. 2: Fecundity VS Body Weight

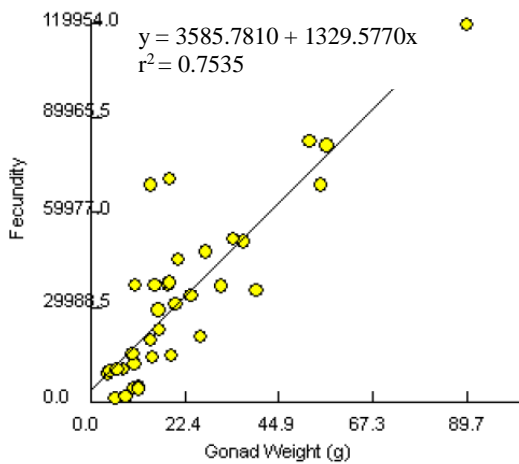


Fig. 3: Fecundity VS Gonad Weight

Plate 1 above shows the fully matured or ripe-running stage of the female gonads of *S.membranaceus* while plate 2 is the male gonad at the maturing stage.

DISCUSSION

The peak (maximum) spawning in *S. membranaceus* was observed in August to October, because more matured gonad stages IV to VI were observed in these months. Araoye (2001) observed peak spawning in *S. schall* to be between June and August. Owolabi (2005) reported a spawning period of *S.membranaceus* in Jebba Lake to be from March to November with the peak between July and September. Lalèyè *et al.*, (2006) observed one spawning season from August to October in *S.schall* and *S.nigrita* from the Ouémé River. Olele and Etim (2011) observed a spawning period of May to August in *S.nigrita* in Onah Lake. Midhat *et al.*, (2012) reported that *S.schall* spawned once from May to August in River Nile at Gizza. Ogbe and Ataguba (2010) also observed that spawning began from June and ended in September in *Mormyrops anguilloides* with principal spawning taking place in August. The period of spawning usually coincides with the period of flooding and highest water volume.

The spawning activities observed in these months (August to October) was the peak of the rainy season being resulting in the expansion of the littoral zone. This is a breeding site for fishes because of availability of spawning substrates and abundant food (Olele, 2010). Food becomes more abundant during the rainy/flood seasons as increased water volume brings about increased food abundance and increased food availability improves fish health condition and thereby enhancing reproduction. Shinkafi and Daneji (2011) reported same, that flood among other things provided expanded habitat and abundant food. That most fishes spawned during this period so as to ensure the survival

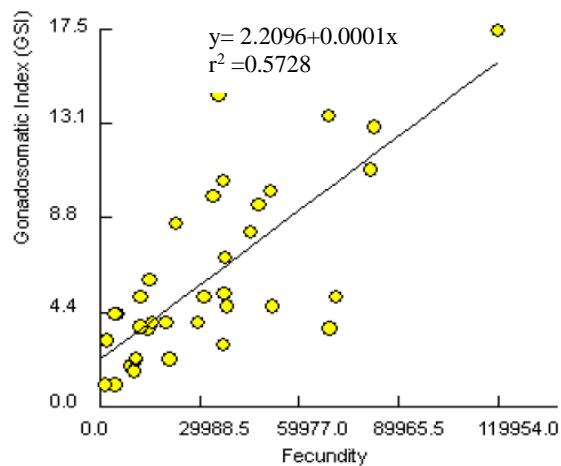


Fig.4: GSI VS Fecundity

of the huge number of the young fish produced, provide reproductive niche and shelter for the juveniles, and consequently ensure reproductive success. Most fishes spawned during this period to maximize the production of recruits. Wootton (1992) stated that a fish should reproduce at that time of the year that will tend to maximize its life time production of offspring. The larval fish must hatch into a world that can provide appropriate food and protection from predators.

The differences in timing observed in the different habitats could be explained on the different environmental and climatic conditions of the habitats.

In this study it was observed that *S.membranaceus* exhibited high fecundity ranging from 1,327 to 119,954 eggs. Willoughby (1974) reported the fecundity of 33,700-179,000 eggs in *S. membranaceus* from Lake Kainji, Nigeria. Owolabi (2005) reported the fecundity of 22,000-267,000 eggs in Jebba Lake, also in Nigeria. From these observations it is clear that *S.membranaceus* has a relatively high fecundity which is an advantage as continued existence of fish depends on the number of eggs hatched and their survival to the adult stage (Olatunde, 1989).

Studies on other species of *Synodontis* reveal that *S.membranaceus* is the most fecund of all the species. In *S.schall*, Araoye (2001) reported the fecundity of 7,910-64,450 eggs from Asa Lake, Ilorin while Lalèyè *et al.*, (2006) reported the fecundity of 1,841-15,076 eggs from Ouémé River, Benin, while Mekkawy and Hassan (2011) reported the fecundity of 1,440-36,920 eggs from the Nile River, Egypt. Willoughby (1974) reported the fecundity of 7,130-73,000 eggs in Lake Kainji, Nigeria while Halim and Guma'a (1989) reported the fecundity of 10,000-90,000 eggs from the White Nile River near Khartoum in Sudan. Olatunde (1989) reported the fecundity of 2,014-13,262 eggs from Zaria while Ofori-Danson (1992) observed the fecundity of 14,000-16,500 eggs from Kpong Head pond, Ghana. In *S.nigrita*, Willoughby, (1974) reported the fecundity of 952-

11,400 eggs from Lake Kainji, Lalèyè *et al.*, (2006) reported the fecundity of 2,647-9,212 eggs in Ouémè River, Benin. Olele and Etim (2011) observed a fecundity of 675-3,642 eggs in Onah Lake while Olojo and Olurin (2012) have reported the fecundity of 216-9,620 eggs from River Osun, Southwest Nigeria.

The great variations in the number of eggs produced by an individual of a certain species were demonstrated by a large number of other tropical fishes by Awachie and Ezenwaji (1981), Lalèyè *et al.*, (1995), Baras and Lalèyè (2003) and many others. Apart from the environmental factors in the different localities, the differences observed may be, in part due to the methods used for fecundity estimation. Iyange and Ezewaji (2004) stated that the discrepancies in fecundity may be as a result of differences in fertility and food availability, disease condition or well-being of the fish and fish body chemistry.

Fecundity in this study revealed that fish of the same length and weight exhibited varied fecundity. This could be attributed to their body chemistry, differences in access to food, differences in their well-being and differences in adaptability to physical and chemical parameters of the water and habitat. Wide fecundity variation among fishes of the same length was reported by Ikomi and Sikoki (2004).

CONCLUSION

The result of this study shows that *S.membranaceus* in River Benue has higher fecundity that is capable of supplementing for losses through mortality and overfishing activities in order to enhance the sustainability of its species throughout the year round.

ACKNOWLEDGMENT

We are most grateful to the laboratory technicians, Mr. Waya, J. I.; Mrs. Shiriki. D.; Mrs. Tyona E.; Mr. Adanu, P., and the Assistants for their support during the practical. We are also grateful to Mr. Atooyough who used to drive us to the market to purchase the fishes and also assisted in the laboratory work, Mr. Atile, J.I. and Miss Shima J.N. who assisted in one way or the other.

REFERENCES

[1] kombo, P. M., Atile, J. I., Adikwu, I. A. and Araoye, P. A. (2011). Morphometric Measurements and growth patterns of four species of the genus *Synodontis* (Cuvier, 1816) from Lower Benue River, Makurdi, Nigeria. *International Journal of Fisheries and Aquaculture*, 3 (15): 263-270.

[2] wachie, J. B. E. and Ezenwaji, H. M. G. (1981b). The importance of *Clarias* species in the

fisheries development of the Anambra Basin, Nigeria. *CIFA Technical Paper*, 8: 212-224.

[3] Bagenal, T. B. and Tesch, F. W. (1978). Methods for assessment of Fish Production I Freshwaters. T. B. Bagenal 9ed.) I. B. P. Handbook No. 33rdedn. *Oxford Blackwell Publications*, 365.

[4] aras, E. and Laèyè, P. (2003). Ecology and Behavior of catfishes, In: Arratia, G., Kapoor, B. G., Chardon, M. and Diogo, R. (eds.) *Cat fishes. Science Publishers, Inc. USA*, 2: 525-579.

[5] eadle, I. C. (1974). The inland waters of Tropical Africa. Longman, 1 – 365.

[6] ishai, H. M. and Abu Gideiri, Y. B. (1965b). Studies on the Biology of genus *Synodontis* at Khartoun II. Food and feeding habits. *Hydrobiologia*, 26: 98-113.

[7] alim, A. I. A. and Guma'a, S. A. (1989). Some aspects of the reproductive biology of *Synodontis schall* (Bloch and Schneider, 1801) from the White Nile near Khartoum. *Hydrobiologia*, 178: 243 - 251.

[8] dodo-Umeh, G. (2003). Fresh water fishes of Nigeria (Taxonomy, Ecological notes, Diet and Utilization). Idodo-Umeh Publishers Ltd. Benin City, Edo State, Nigeria, 11 - 218.

[9] dodo-Umeh, G. (2003). Freshwater fishes of Nigeria (Taxonomy, Ecological notes, Diet and Utilization). *Idodo-Umeh Publishers Ltd.* Benin City, Edo State, Nigeria, 11 - 218.

[10] komi, R.B., Sikoki, F.D. (2003). Studies on the biology of the African lofintera *Brucirus longipinnis ichthyologicaet piscatorial Bol.*

[11] nyang, N. M., Ezenwaji, H. M. G. (2004). Size, Length-weight relationship, Reproduction and Tropical Biology of *Chrysichthys nigrodigitatus* and *Chrysichthy sauratus* (Siluriformes: *Bagridae*) in a Natural West African

[12] esteven, G.L. (1960). Manuals of the field methods in fisheries

[13] agler, K.F.; Bardach, J.E.; Eitter, R.R.; and

- Passimo, D.R.M.(1977). *Ichthyology*. John Wiley and Sons Inc., 506.
- [14] alèyè, P., Chikou, A., Gnohssou, P., Vandewalle, J.C.P. and Teugels, G. (2006). Studies on the Biology of two species of catfish *Synodontis schall* and *Synodontis nigrita* (Ostariophysi: Mochokidae) from the Ouémé River, Bénin. *Belg. Journal. Zoology*, 136 (2): 193 - 201.
- [15] ekkawy, I. A. A. and Hassan, A. A. (2011). Some reproductive Parameters of *Synodontis schall* (Bloch and Schneider,1801) from River Nile, Egypt. *Journal of Fisheries and Aquatic Sciences*, 6 (4): 456-471.
- [16] idhat, A.E.K., Mohammed, M. N., A. and Seham, A. I. (2012). Environmental studies on *Synodontis schall* (Bloch and Schneider, 1801), (Pisces: Mochokidae) in The River Nile at Gizza Sector, Egypt: Biological aspects of Population Dynamics. *Journal of Fisheries and Aquatic*, 7: 104 - 133.
- [17] edeco (1959). Studies and Recommendations: Improvement of Niger and Benue Rivers. Amsterdam. *North Holland Publishing company*, 19 - 27.
- [18] ikolsky, G.V. (1963). The Ecology of Fishes.*Academic press*, New York, 35-41.
- [19] fori-Danson, P. K. (1992). Ecology of some species of catfish *Synodontis* (Pisces: Mochokidae) in the KpongHeadpond Ghana. *Environnemental Biology of Fishes*, 35: 49-61.
- [20] latunde, A. A. (1989). Some Aspects of biology of *Synodontis schall* (Boch and Schneider, 1801) in Zaria, Nigeria. *Journal of Aquatic Sciences*, 4: 49 - 54.
- [21] lele, N. F. and Etim, L. (2011).Some Aspects of the biology of *Synodontis nigrita* (Cuvier and Valenciennes, 1864) in Onah Lake, Asaba, Nigeria.*ARNP Journal of Agricultural and Biological Science*, 6 (1): 56 - 63.
- [22] lojo, E. A. A., Olurin, K. B. and Osikaya, O. J. (2003).Food and feeding habits of *Synodontis nigrita* from the Osun River, SW Nigeria.*NAGA, World Fish Centre Quarterly*, 26: 421 - 424.
- [23] wolabi, O. D. and Omotosho, J. S. (1999).Aspects of the Biology of *Synodontis gambiensis* (Gunther) from Asa Lake, Ilorin, Nigeria. *Nigerian Journal of Pure and Applied Sciences*, 14: 778 - 783.
- [24] wolabi, O. D. (2005). Some Aspects of the Biology of *Synodontis membranaceus* (Goefrey St. Hilaire) in Jebba Lake.Ph D. Thesis, University of Ilorin, Ilorin, Nigeria.*Book of Abstract of Doctoral Theses*, 3: 2005 - 2008.
- [25] eed, W.; Burchark, J.; Hopson, A.M; Jennes, J and Yaro, I. (1967).Fish and Fisheries of Northern Nigeria.*Gaskiya Corporation, Zaria, Northern-Nigeria*, 85-101.
- [26] eid, M.G. and Sydenhan, H.L. (1979).A check-list of Lower Benue River fishes.Ichthyo – geographical review of the Benue River, West-Africa.*Journal of Natural History*, 13: 14-67.
- [27] hinkafi, B. A., Argungu, L. A., and Akanbi, H. S. (2010).Food and Feeding Habits of catfish (*Synodontis nigrita*, Cuvier and Valenciennes) in River Rima, Sokoto, Nigeria.*Nigerian Journal of Basic and Applied Science*, 18 (2): 304 - 307.
- [28] Teugels, G. G. (1996). Taxonomy, Phylogeny and zoogeography of catfishes (Ostariophysi, Siluroidei) an overview, *Aquatic Living Resources* 9: 9-34.
- [29] Olloughby, N. G. (1974).The Ecology of the genus *Synodontis* (Pisces: Siluroidei) in Lake Kainji, Nigeria. Ph.D Thesis, University of Southampton, U.K. In: Olatunde, A. A. (1978). Reproductive cycle and variations in the fecundity of the Family *Scilbeidae* (Osteichthyes: Siluriformes) in Lake Kainji, Nigeria. *Hydrobiologia*, 57 (2): 125 - 142.
- [30] ooten, R. J. (1979). Energy cost of egg production and environmental determinants of fecundity in teleost fishes. *Symp. Zool. Soc. London*, 44: 133 - 159.